



# OSBI Drug Laboratory Training Manual

*Revision #20, Effective Date 06-15-2026*

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## Introduction

The Controlled Substances Laboratory of the Criminalistics Services Division (CSD) of the Oklahoma State Bureau of Investigation (OSBI) is part of an accredited full-service laboratory system responsible for the analysis of samples suspected to contain a controlled dangerous substance. This training manual is intended to provide an analyst with the skills and information needed to perform analysis of submitted samples. Each section of this manual lists a specific goal and the tasks that a trainee should complete in order to achieve this goal. The training will be assessed using an oral examination as well as a competency examination.

At the conclusion of training the trainee should have the following:

1. Knowledge of the principles and practices of forensic marijuana and drug analysis as they relate to the analysis of case material.
2. Knowledge of the theory and application of instrumentation and specialized techniques used to examine marijuana, controlled substances, and non-controlled substances.
3. The skills and ability to perform accurate forensic analysis independently and proficiently, to accurately document the findings of all analysis in accordance with the appropriate policies and procedures, and to accurately generate a report on those findings.

If an analyst has previously passed a mock trial in the OSBI Controlled Substances Unit and has previous experience testifying in court, the analyst may be given an oral examination in lieu of a second mock trial. Two or more Senior Criminalists and the Technical Manager or designee will be present during the mock trial and the "Mock Trial Evaluation Form" will be used for grading. Requirements for passing include a minimum score of a 2 for each section and approval from the Technical Manager. Any score lower than a 2 must have a written explanation for the score.

This training manual can be modified by the Technical Manager for re-training purposes, including an analyst that is returning to the Controlled Substances Unit from another discipline or an analyst that needs retraining in a specific area for remedial reasons.

Once released for casework, it is up to the analyst to seek further training for the maintenance of skills and expertise. The Technical Manager will periodically send out articles for the analyst to read; those articles are to be documented on the Additional Reading/Training form.

The finalized training notebook will be kept by the analyst at the OSBI laboratory. Once training is completed, the training notebook will be scanned and uploaded into the analyst's individual folder on the QA server. Upon termination or transfer to another unit, the training notebook will be scanned, if not already in digital format, and uploaded into the analyst's individual folder on the QA server.



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## Orientation to the OSBI

### Goals

- To ensure the trainee is familiar with forensic science and the different types of forensic services
- To familiarize the trainee with criminal and civil laws that pertain to forensic chemistry
- To familiarize the trainee with the OSBI Drug Quality System
- To introduce the trainee to the OSBI laboratory management systems
- To introduce the trainee to courtroom testimony dynamics and behavior in the courtroom

## General Knowledge of Forensic Science

### Literature

| Date | Literature                                                                                                          |
|------|---------------------------------------------------------------------------------------------------------------------|
|      | Smith, F.P. <b>Overview of Forensic Drug Analysis.</b> <i>Handbook of Forensic Drug Analysis</i> . 2005, pages 1-12 |

## Applicable Criminal and Civil Law and Procedures

### Literature

| Date | Literature                                                                                                                                                                     |
|------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|      | Oklahoma State Statute Title 63, Chapter 2 – Uniform Controlled Dangerous Substances Act, <a href="http://www.oklegislature.gov">www.oklegislature.gov</a>                     |
|      | Federal Code of Regulations Title 21 Part 1308                                                                                                                                 |
|      | Haggerty II, M.D. <b>Confrontation and the Preliminary Hearing.</b> <i>Q &amp; A: The Newsletter of the Criminal Law Section</i> . Vol. 4, Issue 3, May-June 2006, pages 23-31 |
|      | Woodson, M. <b>Relevance and Reliability: What All Expert Testimony Needs.</b> <i>Oklahoma Bar Journal</i> , 79 OBJ 534, March 2008                                            |
|      | Calhoun, M. C. <b>Scientific Evidence in Court: Daubert or Frye, 15 Years Later.</b> <i>Legal Backgrounder</i> , Vol. 23, No. 37, August 22, 2008                              |

### Discussion

| Date | Tasks                                                                                                                              |
|------|------------------------------------------------------------------------------------------------------------------------------------|
|      | Discuss differences in distribution vs. trafficking vs. possession charges                                                         |
|      | Discuss why drugs are scheduled and criteria for different schedules                                                               |
|      | Discuss how drug laws are enacted, by vote of the people & legislative process, including the process of how drugs are controlled. |



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## Drug Quality System Overview

### Literature

| Date | Literature/Tasks                                                       |
|------|------------------------------------------------------------------------|
|      | OSBI <i>Controlled Substances Quality Assurance Manual</i>             |
|      | Review OSBI CSD QM 7.4                                                 |
|      | Review OSBI CSD QM 13, 14.1, 14.2, and 14.3                            |
|      | Read Physical Evidence Quality Procedures Manual 2.1 Evidence Handling |

### Discussion

|  |                            |
|--|----------------------------|
|  | Discuss nonconforming work |
|--|----------------------------|

## Miscellaneous

### Literature

| Date | Literature/Tasks                                                                                                       |
|------|------------------------------------------------------------------------------------------------------------------------|
|      | Review OSBI CSD QMA 2, Evidence Management Requirements, and observe evidence submitting procedures                    |
|      | Review OSBI CSD QMA 3, Evidence Packaging and Sealing Guidelines, and observe evidence sealing and handling procedures |

### Tasks

|  |                                                                                                    |
|--|----------------------------------------------------------------------------------------------------|
|  | The analyst will be shown where to locate the BEAST LIMS system and a brief overview will be given |
|  | The analyst will observe checking out, inventorying and analyzing of a minimum of 5 cases          |
|  | The analyst will be shown where to locate Chemical Inventory and a brief overview will be given    |



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## Testimony & Presentation of Evidence in Court

### Literature

| Date | Literature                                                                                                                                                                                                                                                                     |
|------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|      | Review OSBI Policy 108                                                                                                                                                                                                                                                         |
|      | Shelton Hon., D.E., Barak, G., Kim, Y.S. <b>A Study of Juror Expectation and Demands Concerning Scientific Evidence: Does the “CSI Effect” Exist.</b> <i>Selected Works</i> ( <a href="http://www.works.bepress.com">www.works.bepress.com</a> ). February 2007, pages 331-368 |

### Discussion

| Date | Tasks                                                                                                    |
|------|----------------------------------------------------------------------------------------------------------|
|      | Discuss courtroom testimony and presentation of evidence with trainer                                    |
|      | Discuss bringing and opening evidence in court                                                           |
|      | Discuss requirements for external testing requested by defense, including any accreditation requirements |

### Tasks

|  |                                                                                                     |
|--|-----------------------------------------------------------------------------------------------------|
|  | Review documentation when leaving evidence in court and how to enter the information into the BEAST |
|  | Review a Testimony Review form and Qualified Reviewer form                                          |
|  | Review testimony report and who to send it to                                                       |
|  | Observe a Criminalist from the Controlled Substances Unit giving testimony                          |

Upon signing the approval, the trainee and trainer will review the above information and ensure the trainee has demonstrated knowledge and understanding of the above topics.

### Approval

Trainee \_\_\_\_\_ Date \_\_\_\_\_

Trainer/  
Supervisor \_\_\_\_\_ Date \_\_\_\_\_

Comments \_\_\_\_\_



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## Weights and Measures Utilized in Drug and Marijuana Reports and Balance Scale Calibration and Uncertainty (DR-3 and DR-4)

### Goals

- To establish uniform guidelines in the determination and reporting of weights and volumes of substances submitted for analysis.
- To establish guidelines for procedures to document balance verification, calibration and uncertainty.
- To provide an understanding of the theory of uncertainty of measurement.
- To understand how uncertainty of measurement is calculated and factors that can affect uncertainty.
- To be able to explain uncertainty of measurement in a way a layperson can understand.

### Literature

| Date | Literature                                                                                                                                                                          |
|------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|      | PowerPoint Presentation for Uncertainty of Measurement                                                                                                                              |
|      | Bell, S. A Beginner's Guide to Uncertainty of Measurement. <i>Measurement Good Practice Guide No. 11 (Issue 2)</i> . National Physical Laboratory (PDF: Uncertainty of Measurement) |
|      | Protocol DR-3 (Weights and Measures Utilized in Drug and Marijuana Reports)                                                                                                         |
|      | Protocol DR-4 (Balance/Scale Calibration and Uncertainty)                                                                                                                           |
|      | Protocol DR-4 Attachment 1 (Budget for Calculating Uncertainty of Measurement)                                                                                                      |
|      | Protocol DR-4 Attachment 2 (Controlled Substances Scale Scenarios)                                                                                                                  |
|      | Weighing the Right Way. <i>Guide Book Proper Weighing with Laboratory Balances</i> . Mettler Toledo, 05/2012 (PDF: Uncertainty of Measurement)                                      |
|      | M3003 The Expression of Uncertainty and Confidence in Measurement. United Kingdom Accreditation Service, Edition 2, January 2007                                                    |

### Discussion

| Date | Tasks                                                                                         |
|------|-----------------------------------------------------------------------------------------------|
|      | The definitions for: net weight, gross weight, approximate volume and residue                 |
|      | When gross weights can be utilized                                                            |
|      | The recommended report wording concerning significant digits for the reported weight ranges   |
|      | When a balance is to be checked for proper calibration and when a balance is to be calibrated |
|      | The acceptable operating range of a balance                                                   |



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|                                   |                                                                                                                                 |
|-----------------------------------|---------------------------------------------------------------------------------------------------------------------------------|
|                                   | The proper procedure if a balance is not operating within specified operating range, or if a balance is out of service          |
|                                   | The recommended procedure for checking balances while working cases involving trafficking charges                               |
|                                   | The reason for not reporting weights under a predetermined weight for the different scales                                      |
| <b>Uncertainty of Measurement</b> |                                                                                                                                 |
|                                   | The definition of Uncertainty of Measurement                                                                                    |
|                                   | How uncertainty of measurement is calculated and the factors considered, including budget items vs items not included in budget |
|                                   | How often it is calculated and why it is recalculated                                                                           |
|                                   | Proper report wording of uncertainty and when it is reported                                                                    |
|                                   | Where should uncertainty be added to the report for marijuana cases that include both grams and pounds                          |
| <b>Trafficking levels</b>         |                                                                                                                                 |
|                                   | Trafficking weights for controlled substances (i.e., marijuana, meth, cocaine, cocaine base, etc.)                              |

## Tasks

| Date | Tasks                                                                                              |
|------|----------------------------------------------------------------------------------------------------|
|      | Verify a bench top balance and record on appropriate OSBI DR4 form                                 |
|      | Verify a large capacity scale and record on appropriate OSBI DR4 form                              |
|      | Verify an analytical balance and record on appropriate OSBI DR4 form                               |
|      | Perform 31-day Measurement Assurance Program for bench top balance                                 |
|      | Perform 31-day Measurement Assurance Program for large capacity scale                              |
|      | Perform 31-day Measurement Assurance Program for analytical balance                                |
|      | Demonstrate how to determine the approximate volume of a container using the formula $V=\pi r^2 h$ |

Upon signing the approval, the trainee and trainer will review the above information and ensure the trainee has demonstrated knowledge and understanding of the above topics.

### Approval

Trainee \_\_\_\_\_

Date \_\_\_\_\_

Trainer/  
Supervisor \_\_\_\_\_

Date \_\_\_\_\_

Comments \_\_\_\_\_





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## Thermometers

### Goals

- To establish a guide for the proper reading of thermometers used in the laboratory

### How to Read a Thermometer

Thermometers should be handled carefully because they are tubes of glass filled with either mercury or colored spirits.

Laboratory thermometers should NOT be shaken like the home variety thermometer. To lower the temperature, just cool them in a refrigerator or water/ice bath. Usually, they are either partial or whole immersion thermometers; this means that the bulb may be either partially submerged in a liquid or must be totally submerged in a liquid to accurately register the temperature. Thermometers used in the refrigerators are not to be submerged or placed into any liquid.

Place the thermometer in the material/refrigerator in which the temperature is to be measured. If you are measuring the temperature of a material while it is being heated, make certain that you do not let the thermometer rest on the bottom of the container and that the bulb is submerged in the material itself.

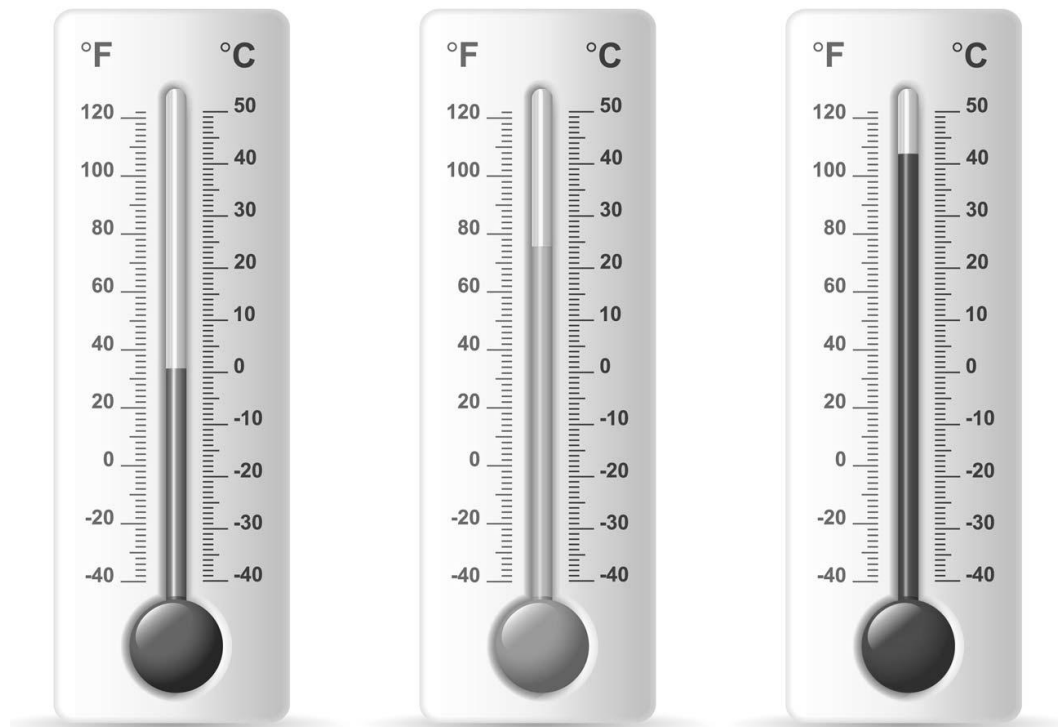
To read the temperature indicated on a thermometer, your eye should be at the level of the liquid in the thermometer. Read the thermometer to the appropriate number of digits. For example, a thermometer on which the heavy or extended lines are marked 10, 20, 30... should be read to the nearest degree.



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First examine the scales below, each degree is divided into smaller divisions. The number of divisions may vary between thermometers, so it is important to look at the scale.



Record the temperature of the three thermometers, to the nearest degree Celsius.

#1 \_\_\_\_\_

#2 \_\_\_\_\_

#3 \_\_\_\_\_

## Literature

| Date | Literature                                                                                          |
|------|-----------------------------------------------------------------------------------------------------|
|      | Read Physical Evidence Quality Procedures Manual 2.4, Evidence Refrigerator and Freezer Maintenance |

## Discussion

| Date | Tasks                                                                                 |
|------|---------------------------------------------------------------------------------------|
|      | What is to be done in the event the refrigerator/freezer is out of temperature range? |
|      | When should monitoring the temperature be performed and how is this documented?       |



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## Tasks

| Date                              | Tasks                                                                                                                                                     |
|-----------------------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------|
| <b>Backup System or Generator</b> |                                                                                                                                                           |
|                                   | Trainer will demonstrate the proper steps to take in the event of a power failure, i.e., use of alternative storage location, backup system or generator. |

## Evaluation of Training

| Date                                                                                   | Tasks                     |
|----------------------------------------------------------------------------------------|---------------------------|
| <b>Demonstrate how to properly read thermometer and document below (5 occurrences)</b> |                           |
|                                                                                        | Temperature: Verified by: |
|                                                                                        | Temperature: Verified by: |
|                                                                                        | Temperature: Verified by: |
|                                                                                        | Temperature: Verified by: |
|                                                                                        | Temperature: Verified by: |

Upon signing the approval, the trainee and trainer will review the above information and ensure the trainee has demonstrated knowledge and understanding of the above topics.

### Approval

Trainee \_\_\_\_\_ Date \_\_\_\_\_

Trainer/  
Supervisor \_\_\_\_\_ Date \_\_\_\_\_

Comments \_\_\_\_\_



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## Cannabis Analysis

This guide is intended to provide a trainee with the necessary skills to perform laboratory analysis of samples suspected to be marijuana, hashish, hashish oil, THC, or THCA-A using a series of examinations. These examinations are stereomicroscopic examination, thin layer chromatography, FTIR, gas chromatography with flame ionization detector and gas chromatography with mass spectral examination.

### Goals

- To become familiar with the legal status of marijuana in Oklahoma.
- To become skilled at the identification of marijuana samples using a series of examinations.

### Literature

| Date | Literature                                                                                                                                                                                     |
|------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|      | Protocol DR-01 (Cannabis Analysis)                                                                                                                                                             |
|      | Protocol DR-01 Attachment 1 – 1% Threshold Testing for Total THC                                                                                                                               |
|      | Protocol DR-01 Attachment 2 – Flowcharts                                                                                                                                                       |
|      | Review Oklahoma Statutes – Title 63, Section 2-101. Specifically, the controlled parts of the marijuana plant.<br>General familiarization with the remainder of the section.                   |
|      | Oklahoma Statutes - Title 22, Section 751. Admission of Laboratory findings.<br>Release of CDS for independent analysis                                                                        |
|      | Nakamura, G.R. <b>Forensic Aspects of Cystolithic Hairs of Cannabis and Other Plants.</b> <i>Journal of the Association of Official Analytical Chemists</i> . Vol. 52, No. 1, 1969, pages 5-16 |
|      | Mechoulam, R. <b>Marihuana Chemistry.</b> <i>Science</i> . Vol. 168, No. 3936, June 5, 1970, pages 1159-1165 (Stop at Biogenesis Section on 3 <sup>rd</sup> page)                              |
|      | <b>Marihuana, Its Identification.</b> US Treasury Department, Bureau of Narcotics, 1948                                                                                                        |
|      | <b>Methods of Analysis.</b> Internal Revenue Service Publication No. 341, Rev. 6-67, page 105                                                                                                  |



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|  |                                                                                                                                                                                                                                                  |
|--|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|  | <b>Lesson Plan #7, Marihuana and THC.</b> <i>Basic Training Program for Forensic Drug Chemists.</i> May 1972, pages 146-157                                                                                                                      |
|  | <b>Analysis of Drugs.</b> <i>DEA Analytical Manual.</i> U.S. Department of Justice, pages 165-168                                                                                                                                                |
|  | Coutts, R.T. & Jones, G.R. <b>A Comparative Analysis of Cannabis Material.</b> <i>Journal of Forensic Science.</i> Vol. 24, No. 2, 1979, pages 291-302.                                                                                          |
|  | Small, E. <b>American Law and the Species Problem in Cannabis.</b> <i>Microgram.</i> Vol. VII, No. 11, November 1974, pages 131-132.                                                                                                             |
|  | Nakamura, G.R. and Thornton, J.I. <b>The Forensic Identification of Marihuana: Some Questions and Answers.</b> <i>Journal of Police Science and Administration.</i> Vol. I, No. I, 1973, pages 102-112                                           |
|  | Zimmerman, Miles C. <b>Marijuana Analysis: Winters V. State.</b> <i>OSBI Legal Update.</i> Index Tab: Drugs, Control No. 07-77-02, March 24, 1977                                                                                                |
|  | <b>Marihuana.</b> <i>Basic Training Program for Forensic Drug Chemists.</i> 2 <sup>nd</sup> Edition, DEA, pages 6-25 to 6-44                                                                                                                     |
|  | Johnson, Donald W. <b>Hashish Oil.</b> <i>DEA Laboratory Notes.</i> No. 58, May 1973                                                                                                                                                             |
|  | <b>Cannabis.</b> <i>The Drug Chromatographer.</i> Alltech-Applied Science, Vol. 3, Number 1, 1986, pages 1-3                                                                                                                                     |
|  | <b>Recommended Methods for the Identification and Analysis of Cannabis and Cannabis Products.</b> United Nations Office on Drugs and Crime, 2009                                                                                                 |
|  | Warner, M.L., Alford, I., Lawrence, D.M., Kohl, A.C., Williams, S.J., Yeatman, D.T. <b>Comparative Analysis of Freshly Harvested Cannabis Plant Weight and Dried Cannabis Plant Weight.</b> <i>Forensic Chemistry.</i> Vol. 3, 2017, pages 52-57 |
|  | Hughes, R. B. M. S. and Kessler, R. R., M.S. <b>Increased Safety and Specificity in the Thin-Layer Chromatographic Identification of Marihuana.</b> <i>Journal of Forensic Sciences.</i> Vol. 24, March 19, 1979, pages 842-846                  |
|  | Guy, B. <b>The Identification of Cannabis by Thin Layer Chromatography.</b> <i>Microgram.</i> Vol. XVII, No. 5, May 1984, pages 78-80                                                                                                            |



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## Discussion

| Date                              | Tasks                                                                                                                |
|-----------------------------------|----------------------------------------------------------------------------------------------------------------------|
| <b>Microscopic Examination</b>    |                                                                                                                      |
|                                   | Microscopic characteristics of marijuana                                                                             |
|                                   | Requirement for a positive examination                                                                               |
|                                   | Discuss what a typical marijuana leaf looks like, including vein structure                                           |
| <b>Medical Marijuana and Hemp</b> |                                                                                                                      |
|                                   | Differences between hemp and marijuana                                                                               |
|                                   | Are there differences in analysis of medical marijuana and illegal marijuana                                         |
|                                   | Discuss THC vs THCA                                                                                                  |
| <b>Analysis</b>                   |                                                                                                                      |
|                                   | Proper method for analysis of seeds                                                                                  |
|                                   | Some of the possible indicators that a second controlled dangerous substance may be present in a marijuana submittal |
|                                   | How to analyze a green leafy sample                                                                                  |
|                                   | How to analyze a wax or oil-like substance                                                                           |
|                                   | Articulate how to analyze a food/drink product                                                                       |
|                                   | The proper procedure for TLC with a sample suspected to contain delta 9 tetrahydrocannabinol                         |
|                                   | Articulate interpretation criteria of TLC plate results, such as height of a sample/standard, and color of spots     |
|                                   | When a TLC plate needs to be rejected                                                                                |
|                                   | When should a sample use the 1% protocol                                                                             |

## Tasks

| Date                           | Tasks                                                               |
|--------------------------------|---------------------------------------------------------------------|
| <b>Microscopic Examination</b> |                                                                     |
|                                | Demonstrate use of a stereomicroscope including magnification range |
| <b>Analysis</b>                |                                                                     |
|                                | Demonstrate the preparation and analysis of a sample using TLC      |
|                                | Demonstrate how to extract a cannabis sample using BSTFA            |
|                                | Prepare TLC Reagent                                                 |
|                                | Demonstrate how to use the calibrated pipettes                      |
|                                | Demonstrate how to prepare a sample for the 1% protocol             |



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Upon signing the approval, the trainee and trainer will review the above information and ensure the trainee has demonstrated knowledge and understanding of the above topics.

## **Approval**

Trainee \_\_\_\_\_ Date \_\_\_\_\_

Trainer/  
Supervisor \_\_\_\_\_ Date \_\_\_\_\_

Comments \_\_\_\_\_



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## Drug Literature

This guide is intended to provide a trainee with the necessary skills to perform laboratory analysis of samples suspected to contain a controlled substance. This analysis will involve the use of various instruments and laboratory techniques to reach conclusions on the identification of a substance.

### Goals

- To provide a general understanding of controlled substances

### Literature

| Date | Literature                                                                                                                                                              |
|------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|      | Review Oklahoma Statutes – Title 63, Section 2-101, 2-201 thru 2-212, 2-321, 2-407.1, 2-414, 2-415                                                                      |
|      | Bell, S. What is a Drug. <i>Forensic Chemistry</i> . pages 213-231, 234-239                                                                                             |
|      | Drugs of Abuse. US Department of Justice, Drug Enforcement Agency, 2011                                                                                                 |
|      | Gahlinger, P.M. <i>Illegal Drugs, A Complete Guide to Their History, Chemistry, Use and Abuse</i> . pages 232-236 (PDF: Cathinone)                                      |
|      | Synthetic Cathinones (“Bath Salts”). National Institute on Drug Abuse, <a href="http://www.drugabuse.gov">www.drugabuse.gov</a> , 01/10/2013                            |
|      | <i>Drug Identification Bible</i> . 2022/2023, MDMA, pages 646-652                                                                                                       |
|      | Gahlinger, P.M. <i>Illegal Drugs, A Complete Guide to Their History, Chemistry, Use and Abuse</i> . pages 264-275 (PDF: Hallucinogens (DMT, Bufotenine and Psilocybin)) |
|      | Gahlinger, P.M. <i>Illegal Drugs, A Complete Guide to Their History, Chemistry, Use and Abuse</i> . pages 224-228 (PDF: Barbiturates)                                   |
|      | <i>Drug Identification Bible</i> . 2022/2023, Anabolic Steroids, pages 581-584                                                                                          |
|      | <i>Drug Identification Bible</i> . 2022/2023, Heroin, pages 609-623                                                                                                     |
|      | Recommended Methods for Testing Opium, Morphine and Heroin. United Nations Office on Drugs and Crime, 1998 (Modified PDF version)                                       |
|      | <i>Drug Identification Bible</i> . 2022/2023, Fentanyl, pages 600-605                                                                                                   |
|      | <i>Drug Identification Bible</i> . 2022/2023, PCP, pages 653-656                                                                                                        |
|      | <i>Drug Identification Bible</i> . 2022/2023, Ketamine, pages 624-626                                                                                                   |
|      | <i>Drug Identification Bible</i> . 2022/2023, Amphetamine/Methamphetamine, pages 567-580                                                                                |





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|--|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|  | Kelly, B.C. Legally Tripping: A Qualitative Profile of <i>Salvia Divinorum</i> Use Among Young Adults. <i>Journal of Psychoactive Drugs</i> . Vol. 43 (1), 2011, pages: 46-54                                                                        |
|  | Harris, D. et al. GC-MS Differentiation of Three Synthetic Cannabinoid Positional Isomers: JWH-250, JWH-302 and JWH-201. <i>Journal of the Clandestine Laboratory Investigating Chemists Association</i> . Vol. 21, No. 4, October 2011, pages 23-32 |
|  | Protocol DR-103 (Classification of Synthetic Cannabinoids)                                                                                                                                                                                           |

## Discussion

| Date | Tasks                                        |
|------|----------------------------------------------|
|      | Convert milligrams to grams                  |
|      | Vicks inhaler and what substance it contains |

Upon signing the approval, the trainee and trainer will review the above information and ensure the trainee has demonstrated knowledge and understanding of the above topics.

### Approval

Trainee \_\_\_\_\_ Date \_\_\_\_\_

Trainer/  
Supervisor \_\_\_\_\_ Date \_\_\_\_\_

Comments \_\_\_\_\_



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## Pharmaceutical Identification by Literary Reference (DR-5)

### Goals

- To establish guidelines when using visual examinations and comparison to a literary reference as the presumptive examinations for tablet and capsule exhibits.

### Literature

| Date |  |
|------|--|
|      |  |
|      |  |
|      |  |

### Discussion

| Date | Tasks                                                                            |
|------|----------------------------------------------------------------------------------|
|      | When a literary reference is sufficient and when a conclusive analysis is needed |
|      | The procedure when a suspected tablet or capsule was clandestinely manufactured  |
|      | The different resources that can be used for a literary reference                |

### Tasks

| Look up at least 5 tablet/capsules using the references in DR-05 |                    |           |                                           |
|------------------------------------------------------------------|--------------------|-----------|-------------------------------------------|
| Date                                                             | Tablet Description | Reference | Result (Identification and Concentration) |
|                                                                  |                    |           |                                           |
|                                                                  |                    |           |                                           |
|                                                                  |                    |           |                                           |
|                                                                  |                    |           |                                           |
|                                                                  |                    |           |                                           |



# OSBI Drug Laboratory Training Manual

*Revision #20, Effective Date 06-15-2026*

Upon signing the approval, the trainee and trainer will review the above information and ensure the trainee has demonstrated knowledge and understanding of the above topics.

## **Approval**

Trainee \_\_\_\_\_ Date \_\_\_\_\_

Trainer/  
Supervisor \_\_\_\_\_ Date \_\_\_\_\_

Comments \_\_\_\_\_



# OSBI Drug Laboratory Training Manual

Revision #20, Effective Date 06-15-2026

## Training Procedures for Extractions & Handling

### Goals

- To familiarize the analyst with extraction and handling procedures commonly used to prepare samples for analysis using gas chromatography (GC) and gas chromatography-mass spectrometry (GC/MS).

### Literature

| Date | Literature                                                                                                                                                                                                                                                               |
|------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|      | Review SDS sheets on appropriate chemicals used, such Sodium Hydroxide, Hydrochloric Acid, Sodium Bicarbonate, Chloroform, Isopropanol, Hexanes, Methanol, etc.                                                                                                          |
|      | Protocol DR-110 (Extractions) and Appendix C: Extraction Data                                                                                                                                                                                                            |
|      | Review extraction references in the Training folder<br><a href="#">\\VM-FSC-FILES\Common\DrugLab\1 Technical Manager\1 Training\Extraction References</a><br><i>*These are drugs that aren't seen on a regular basis but should be reviewed in case they are needed.</i> |
|      | NIJ Fingerprint Sourcebook, Sections 7.1.5 Evidence Handling and 7.1.6 Packaging                                                                                                                                                                                         |

### Discussion

| Date | Tasks                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                |
|------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|      | Which solvent is a good, all-around solvent for most drugs <ul style="list-style-type: none"><li>Differences in solubility of different drugs</li></ul>                                                                                                                                                                                                                                                                                                                                                              |
|      | When other extractions should be used <ul style="list-style-type: none"><li>The extraction produces a GC analysis cluttered with peaks</li><li>The extraction produces a GC analysis where a secondary peak (i.e., acetaminophen) dwarfs the peak of interest</li><li>The extraction produces a GC analysis where poor separation or broad peaks occur</li><li>Presumptive color tests indicate amphetamine or methamphetamine</li><li>Literary reference indicates a substance that needs to be extracted</li></ul> |
|      | Why drugs need to be basic extracted <ul style="list-style-type: none"><li>Methamphetamine/Amphetamine; Ephedrine/Pseudoephedrine; Phenethylamines</li></ul>                                                                                                                                                                                                                                                                                                                                                         |
|      | Which drugs need to be acid extracted                                                                                                                                                                                                                                                                                                                                                                                                                                                                                |
|      | When to use a back extraction                                                                                                                                                                                                                                                                                                                                                                                                                                                                                        |
|      | Definition of amphoteric and know which drugs exhibit this property                                                                                                                                                                                                                                                                                                                                                                                                                                                  |



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|  |                                                                                                                                                                                   |
|--|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|  | Which solvent systems should be used with morphine and hydromorphone                                                                                                              |
|  | What to do when tablets form an emulsion                                                                                                                                          |
|  | Which substances experience rapid breakdown in solution, and what to do when this occurs (i.e., oxymetholone)                                                                     |
|  | Describe what is to occur if told a sample needs to be placed on the instrument immediately                                                                                       |
|  | Explain how to mix 150 ml of a 2:5:3 solution of A:B:C                                                                                                                            |
|  | Discuss proper packaging for fentanyl and liquids                                                                                                                                 |
|  | Discuss working fentanyl on the GC/IRD <ul style="list-style-type: none"> <li>• What samples are suitable for the GC/IRD</li> <li>• Fentanyl and isomers on the GC/IRD</li> </ul> |

## Tasks

| Date                        | Tasks                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                             |
|-----------------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| <b>Solution Preparation</b> |                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                   |
|                             | Prepare 0.45N NaOH/DI water solution                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                              |
|                             | Prepare 10% HCl/DI water solution. H <sub>2</sub> SO <sub>4</sub> can be substituted for HCl                                                                                                                                                                                                                                                                                                                                                                                                                                                                                      |
|                             | Prepare a saturated sodium bicarbonate solution. H <sub>2</sub> SO <sub>4</sub> can be substituted for HCl                                                                                                                                                                                                                                                                                                                                                                                                                                                                        |
| <b>Extractions</b>          |                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                   |
|                             | Determine the pH of a solution                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                    |
|                             | Demonstrate the procedure for performing a basic extraction <ol style="list-style-type: none"> <li>1. Add 0.45N NaOH or another appropriate basic solution to the sample</li> <li>2. Add appropriate amount of chloroform or hexanes</li> <li>3. Mix thoroughly and centrifuge if necessary <ul style="list-style-type: none"> <li>• Chloroform is preferred over hexanes since it can solubilize a greater number of drugs</li> </ul> </li> <li>4. When utilizing this extraction procedure chloroform will form the bottom layer and hexanes will form the top layer</li> </ol> |
|                             | Demonstrate the procedure for performing an acidic extraction <ol style="list-style-type: none"> <li>1. Add 5-10 drops of 10% HCl or H<sub>2</sub>SO<sub>4</sub> to the sample</li> <li>2. Add appropriate amount of chloroform</li> <li>3. Mix thoroughly and centrifuge if necessary</li> </ol>                                                                                                                                                                                                                                                                                 |
|                             | Demonstrate the procedure for a back extraction <ol style="list-style-type: none"> <li>1. Add 1 milliliter of DI water to a sample</li> <li>2. Add 5-10 drops of 10% HCl solution to the sample</li> <li>3. Mix well and centrifuge if necessary</li> <li>4. Remove the aqueous layer and place in another culture tube</li> <li>5. Make the aqueous layer basic by adding 0.45N NaOH solution</li> </ol>                                                                                                                                                                         |



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|  |                                                                                                                                                                                                                                                         |
|--|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|  | <ol style="list-style-type: none"><li>6. Verify the pH of the aqueous solution has converted from an acid to a base</li><li>7. Once basic, add an appropriate solvent (chloroform or hexanes)</li><li>8. Mix well and centrifuge if necessary</li></ol> |
|--|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|

| Date                   | Demonstrate                                                                                                                                                 |
|------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------|
| <b>Sample Handling</b> |                                                                                                                                                             |
|                        | Demonstrate how to label evidence and test tubes/vials                                                                                                      |
|                        | Demonstrate how to dilute and concentrate a sample                                                                                                          |
|                        | Safely remove clean syringe with needle (previously prepared by trainer) from sharps container, rinse with a solvent, recap and replace back into container |
|                        | Safely remove razor or knife from sharps container (previously prepared by trainer), sample (i.e., with swab) and replace back into container.              |
|                        | Safely remove broken glass from envelope (previously prepared by trainer), sample residue from broken glass, and place into an appropriate sharps container |
|                        | Demonstrate handling, marking, & sampling of evidence to be forwarded to Latent Evidence Unit, with minimal risk of damaging potential latent prints        |

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## Approval

Trainee \_\_\_\_\_ Date \_\_\_\_\_

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# OSBI Drug Laboratory Training Manual

Revision #20, Effective Date 06-15-2026

## Identification of Gamma-Hydroxybutyric Acid through Derivatization (DR-7)

### Goals

- To establish guidelines for the identification of Gamma-Hydroxybutyric Acid (GHB) through derivatization with bis-(trimethylsilyl) trifluoroacetamide (BSTFA) and analysis using gas chromatography (GC) and gas chromatography mass spectrometry (GC/MS).

### Literature

| Date | Literature                                                                                                                                                                                                                                                                 |
|------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|      | Protocol DR-7 (Identification of Gamma-Hydroxybutyric Acid through Derivatization)                                                                                                                                                                                         |
|      | SDS sheets for BSTFA and acetonitrile                                                                                                                                                                                                                                      |
|      | <i>Drug Identification Bible</i> . 2022/2023, GHB, pages 606-608                                                                                                                                                                                                           |
|      | Kilpatrick, G. A. <b>GHB</b> . State Police-San Francisco.                                                                                                                                                                                                                 |
|      | Bell, S. <b>Derivatization</b> . <i>Forensic Chemistry</i> . pages 203-205                                                                                                                                                                                                 |
|      | Bommarito, C. <b>Analytical Profile of Gamma-Hydroxybutyric Acid (GHB)</b> . <i>Journal of the Clandestine Laboratory Investigating Chemists Association</i> . Vol. 3, No. 3, July 1993, pages 10-12                                                                       |
|      | Pearson, J.R., Reid, E.F., & Rowe, J.E. <b>The Preparation of <math>\gamma</math>-Butyrolactone from Readily Available Starting Materials</b> . <i>Journal of the Clandestine Laboratory Investigating Chemists Association</i> . Vol. 19, No. 1, January 2009, pages 8-13 |

### Discussion

| Date | Tasks                                                                                         |
|------|-----------------------------------------------------------------------------------------------|
|      | The theory of derivatization and why it is performed in the differentiation of GHB and GBL    |
|      | How to recognize when GHB may be present in a sample                                          |
|      | The reason for washing a liquid sample suspected of containing GHB and/or GBL with chloroform |
|      | How heat or pH may affect a sample containing GHB and/or GBL                                  |
|      | Proper reporting results for both dry and liquid samples                                      |



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Revision #20, Effective Date 06-15-2026

## Tasks

| Date | Tasks (If samples are available)                                  |
|------|-------------------------------------------------------------------|
|      | Demonstrate derivatization procedure on a dry sample              |
|      | Demonstrate derivatization procedure on a liquid sample           |
|      | Demonstrate ability to identify GBL if present in a liquid sample |

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Trainer/  
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# OSBI Drug Laboratory Training Manual

Revision #20, Effective Date 06-15-2026

## Training Procedures for Color Tests (DR-10, DR-11, DR-13)

### Goals

- To establish guidelines for preliminary screening tests that respond to particular functional groups on substances causing characteristic color changes. These examinations can give the analyst a basis as to which extractions and/or examinations are necessary for further conclusive instrumental analysis.

### Literature

| Date | Literature                                                            |
|------|-----------------------------------------------------------------------|
|      | Protocol DR-10 (Color Tests: Marquis)                                 |
|      | Protocol DR-11 (Color Tests: Cobalt Thiocyanate)                      |
|      | Protocol DR-13 (Color Tests: Bates)                                   |
|      | Clarke's Analysis of Drugs and Poisons, Third Edition, pages 279 -300 |

### Discussion

| Date | Task                                                                                                  |
|------|-------------------------------------------------------------------------------------------------------|
|      | The specificity of color tests and their role in drug analysis                                        |
|      | Where all color tests are to be performed                                                             |
|      | Reasons why a color test may be negative for a compound even when the compound is present in a sample |
|      | Indications of when a reagent may need to be discarded and new reagent made                           |
|      | The procedure for performing a negative control and when it is necessary                              |
|      | The procedure for performing a positive control and when it is necessary                              |



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## Tasks

| Tasks                                                                                       |              |                       |             |
|---------------------------------------------------------------------------------------------|--------------|-----------------------|-------------|
|                                                                                             | Marquis Test | Cobalt<br>Thiocyanate | Bates' Test |
| Demonstrate the procedure for making the reagent                                            |              |                       |             |
| Demonstrate the quality control verification and documentation procedure                    |              |                       |             |
| Demonstrate the procedure for performing the test                                           |              |                       |             |
| Based on the observed results of the color test, articulate a suitable extraction procedure |              |                       |             |

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Supervisor \_\_\_\_\_

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# OSBI Drug Laboratory Training Manual

Revision #20, Effective Date 06-15-2026

## Cocaine Free-base Determination by Hexane Solubility (DR-21)

### Goals

- To establish a knowledge of cocaine and cocaine base.
- To establish guidelines to differentiate cocaine hydrochloride from cocaine free base based on their solubility in hexanes.

### Literature

| Date | Literature                                                                                                                                 |
|------|--------------------------------------------------------------------------------------------------------------------------------------------|
|      | Protocol DR-21 (Cocaine Free Base Determination by Hexane Solubility)                                                                      |
|      | Gahlinger, P.M. <i>Illegal Drugs, A Complete Guide to Their History, Chemistry, Use and Abuse</i> . pages 240-254 (PDF: Cocaine)           |
|      | <b>Cocaine</b> . <i>NIDA InfoFacts</i> . <a href="http://www.drugabuse.gov">www.drugabuse.gov</a> , 01/10/2013                             |
|      | <b>Recommended Methods for Identification and Analysis of Cocaine in Seized Materials</b> . United Nations Office on Drugs and Crime, 2012 |
|      | Crack Cocaine Recipe, <a href="http://www.hyperreal.org">www.hyperreal.org</a> , 1992                                                      |
|      | <i>Drug Identification Bible</i> . 2022/2023, Cocaine, pages 585-599                                                                       |

### Discussion

| Date | Tasks                                                                            |
|------|----------------------------------------------------------------------------------|
|      | Why cocaine hydrochloride and cocaine base need to be differentiated             |
|      | What color tests can be used to indicate the presence of cocaine or cocaine base |
|      | The procedure if a sample is negative for cocaine base                           |
|      | The difference between cocaine base and cocaine HCl                              |



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## Tasks

| Date | Tasks                                                                                                                                    |
|------|------------------------------------------------------------------------------------------------------------------------------------------|
|      | Demonstrate the difference in solubility of cocaine hydrochloride and cocaine base using both methanol and hexanes                       |
|      | Demonstrate the difference in reactivity of cocaine hydrochloride and cocaine base with the Cobalt Thiocyanate color test and Bates Test |

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Supervisor \_\_\_\_\_ Date \_\_\_\_\_

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# OSBI Drug Laboratory Training Manual

Revision #20, Effective Date 06-15-2026

## Identification of Lysergic Acid Diethylamide (DR-101)

### Goals

- To establish a knowledge of Lysergic Acid Diethylamide (LSD).
- To familiarize the analyst with procedures used in the identification of LSD in different forms.

### Literature

| Date | Literature                                                                                                                     |
|------|--------------------------------------------------------------------------------------------------------------------------------|
|      | Protocol DR-101 (Identification of Lysergic Acid Diethylamide)                                                                 |
|      | <i>Drug Identification Bible</i> . 2022/2023, LSD, pages 627-630                                                               |
|      | <b>Recommended Methods for Testing Lysergide (LSD)</b> . United Nations Office on Drugs and Crime, 1989 (Modified PDF version) |
|      | Smith, F., <i>Handbook for Drug Analysis</i> . 2005, pages 186-187                                                             |

### Discussion

| Date | Tasks                                                                 |
|------|-----------------------------------------------------------------------|
|      | Presumptive test for LSD on a sample                                  |
|      | Which GC/MS methods would be used for an LSD sample and reagent blank |
|      | Explain how to sample blotter paper                                   |
|      | What report wording is used when analyzing suspected LSD              |

### Tasks

| Date | Tasks                                                                                                               |
|------|---------------------------------------------------------------------------------------------------------------------|
|      | Analyze LSD, LAMPA and a mixture of LSD & LAMPA to demonstrate the differences of the compounds on the GC and GC/MS |

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# OSBI Drug Laboratory Training Manual

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## Gas Chromatography Analysis (Flame Ionizing Detector) (DR-30)

### Goals

- To gain knowledge of the theory of gas chromatography and how to use it as a non-confirmatory test in the analysis of submitted samples.
- To establish guidelines for the gas chromatograph maintenance.
- To demonstrate a gas chromatograph is working properly by using quality assurance and quality control methods.
- The analyst will learn how to interpret data from this type of analysis.

### Literature

| Date | Literature                                                               |
|------|--------------------------------------------------------------------------|
|      | Protocol DR-30 (Gas Chromatography Analysis (flame ionization detector)) |
|      | Basic Operation of the Gas Chromatograph (Appendix I of Training Manual) |
|      | Bell, S. <i>Forensic Chemistry</i> . pages 192-200 (PDF: GC and MS)      |

### Discussion

| Date                                | Tasks                                                                                                                         |
|-------------------------------------|-------------------------------------------------------------------------------------------------------------------------------|
| <b>Theory of Gas Chromatography</b> |                                                                                                                               |
|                                     | The injection port                                                                                                            |
|                                     | Expansion volumes and how to determine the appropriate injection volume                                                       |
|                                     | Megabore columns used in the gas chromatograph (DB-1 and DB-50)                                                               |
|                                     | The flame ionizing detector                                                                                                   |
|                                     | Split ratios and why it is used with the instrument                                                                           |
|                                     | The different methods used for analysis, such as Drug1, Extend1 and Method1 and/or any other methods being used for analysis  |
|                                     | Make-up gas                                                                                                                   |
|                                     | Retention time and how it applies to gas chromatograph analysis                                                               |
| <b>Controls</b>                     |                                                                                                                               |
|                                     | When standard ladders are to be run                                                                                           |
|                                     | What is the negative control and when should it be ran                                                                        |
|                                     | What is the positive control and when should it be ran                                                                        |
|                                     | When methanol blanks should be run                                                                                            |
|                                     | The proper corrective procedure if the retention time of the cocaine standard exceeds plus or minus 2% of the standard ladder |
|                                     | What, if any, type of extraneous peaks is allowed in cocaine standard                                                         |
|                                     | What constitutes contamination/carryover in methanol blanks                                                                   |
|                                     | What are the acceptable levels or sizes of contamination/carryover peaks allowed                                              |



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|              |                                                                                                                                                                                                               |
|--------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|              | The proper corrective procedure if contamination occurs during methanol blank runs                                                                                                                            |
|              | The proper corrective procedure if the instrument fails to produce a satisfactory chromatogram for the cocaine standard                                                                                       |
| <b>Misc.</b> |                                                                                                                                                                                                               |
|              | What is the temperature programming and where can it be found                                                                                                                                                 |
|              | When must a standard be run on the gas chromatograph                                                                                                                                                          |
|              | The maximum number of days between running a standard and a sample on the gas chromatograph                                                                                                                   |
|              | When changing of the liner and septa is required                                                                                                                                                              |
|              | When changing of the gold seal is required                                                                                                                                                                    |
|              | When cleaning of the flame ionization detector is required                                                                                                                                                    |
|              | When cleaning of the injection port is required                                                                                                                                                               |
|              | What conditions and/or situations destroy the stationary phase of a GC column                                                                                                                                 |
|              | Articulate reasons that GC data may be "rejected"                                                                                                                                                             |
|              | What happens to sample after analysis through instrument? <ul style="list-style-type: none"> <li>• Where does the waste from split vent go?</li> <li>• Where does sample go after leaving the jet?</li> </ul> |

## Tasks

| Date                         | Tasks                                                                                                                                                       |
|------------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------|
| <b>Maintenance</b>           |                                                                                                                                                             |
|                              | Change the liner and septum and reset macro counts                                                                                                          |
|                              | Change the gold seal, clean the injector port and septa nut and reset macro counts                                                                          |
|                              | Clean the flame ionization detector                                                                                                                         |
|                              | Change a column when necessary                                                                                                                              |
|                              | Change the split vent filter and line                                                                                                                       |
|                              | Demonstrate syringe replacement                                                                                                                             |
|                              | Demonstrate proper wash and waste bottle volumes                                                                                                            |
|                              | Extract standard ladder, run on GC and update macros                                                                                                        |
|                              | Record any maintenance performed on the proper maintenance log                                                                                              |
| <b>Use of the Instrument</b> |                                                                                                                                                             |
|                              | Properly prepare a sample in an auto-sampler vial                                                                                                           |
|                              | Properly load a sample for analysis on the gas chromatograph, including entering information in sequence log                                                |
|                              | Demonstrate the proper use of controls: <ul style="list-style-type: none"> <li>• MeOH blank</li> <li>• Cocaine standard</li> <li>• Reagent blank</li> </ul> |



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| Spectral Interpretation |                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                              |
|-------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|                         | <p>The analyst will review instrumental data and discuss with the trainer what is and is not acceptable for casework analysis</p> <ul style="list-style-type: none"><li>• Samples outside of the 2% retention time window</li><li>• Demonstrate calculating the 2% window based on the retention time of a standard</li><li>• Peak separation</li><li>• Acceptable/unacceptable chromatography</li><li>• How to integrate a peak in data analysis</li></ul> <p><i>* See Training folder for examples of instrumental data that might need more work</i><br/><a href="#">\\VM-FSC-FILES\Common\DrugLab\1 Technical Manager\1 Training</a></p> |
|                         | Properly interpret the data obtained and explain how to apply this data if further analysis is required                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                      |
|                         | Demonstrate the proper reporting of results available from gas chromatographic data                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                          |

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## Approval

Trainee \_\_\_\_\_ Date \_\_\_\_\_

Trainer/  
Supervisor \_\_\_\_\_ Date \_\_\_\_\_

Comments \_\_\_\_\_





# OSBI Drug Laboratory Training Manual

Revision #20, Effective Date 06-15-2026

## Analysis of Mushrooms to Determine the Presence of Psilocyn or Psilocybin (DR-45)

### Goals

- To establish guidelines for the differentiation of psilocyn and psilocybin for identification.

### Literature

| Date | Literature                                                                                                                                                                                 |
|------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|      | Protocol DR-45 (Analysis of Mushrooms to Determine Presence of Psilocyn or Psilocybin)                                                                                                     |
|      | <i>Drug Identification Bible</i> . 2022/2023, Peyote & Psilocybin Mushrooms, pages 657-663                                                                                                 |
|      | <b>Recommended Methods for Testing Peyote Cactus (Mescal Buttons)/Mescaline and Psilocybe Mushrooms/Psilocybin</b> . United Nations Office on Drugs and Crime, 1989 (Modified PDF version) |

### Discussion

| Date                                         | Tasks                                                                                                                                    |
|----------------------------------------------|------------------------------------------------------------------------------------------------------------------------------------------|
|                                              | Why psilocyn and psilocybin need to be differentiated                                                                                    |
|                                              | What happens to psilocyn and psilocybin when injected into the gas chromatograph in methanol                                             |
|                                              | The different methods that can be used to differentiate psilocyn and psilocybin                                                          |
| <b>Thin Layer Chromatography Examination</b> |                                                                                                                                          |
|                                              | Articulate proper procedure and specificity for TLC with a sample suspected to contain psilocyn or psilocybin                            |
|                                              | Articulate familiarization with Rf value                                                                                                 |
|                                              | Articulate other procedure(s) that has to be performed in conjunction with TLC with a sample suspected to contain psilocyn or psilocybin |
| <b>Derivatization of Sample</b>              |                                                                                                                                          |
|                                              | Articulate proper procedure and specificity for derivatization with a sample suspected to contain psilocyn or psilocybin                 |



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## Tasks

| Date                                         | Tasks                                                                                                                  |
|----------------------------------------------|------------------------------------------------------------------------------------------------------------------------|
| <b>Thin Layer Chromatography Examination</b> |                                                                                                                        |
|                                              | Prepare Mushroom TLC Reagent                                                                                           |
|                                              | Demonstrate the preparation and analysis of a sample using TLC                                                         |
|                                              | Demonstrate the two ways for visualization of a TLC plate for a sample suspected to contain psilocyn and/or psilocybin |
|                                              | Establish interpretation criteria of TLC test results, such as height of a sample/standard, and color of spots         |
| <b>Derivatization of Sample</b>              |                                                                                                                        |
|                                              | Demonstrate derivatization of a sample suspected to contain psilocyn and/or psilocybin                                 |

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Trainee \_\_\_\_\_ Date \_\_\_\_\_

Trainer/  
Supervisor \_\_\_\_\_ Date \_\_\_\_\_

Comments \_\_\_\_\_



# OSBI Drug Laboratory Training Manual

Revision #20, Effective Date 06-15-2026

## PDF Examination Documentation Procedure (DR-50)

### Goals

- To establish guidelines for generating, storing, transferring and attaching instrumental data into the BEAST Image Vault.
- The analyst will learn the security and tracking features associated with this process.

### Literature

| Date | Tasks                                                    |
|------|----------------------------------------------------------|
|      | Protocol DR-50 (PDF Examination Documentation Procedure) |
|      | Review DRQM-11 Examination Documentation                 |

### Discussion

| Date | Tasks                                                                                                                 |
|------|-----------------------------------------------------------------------------------------------------------------------|
|      | Articulate the security and tracking features associated with this process (creation, merging, and uploading of PDFs) |
|      | What the validation code is and where it comes from                                                                   |
|      | Articulate the manner for archiving PDFs                                                                              |

### Tasks

| Date | Tasks                                                                                          |
|------|------------------------------------------------------------------------------------------------|
|      | Set up PDF folders on the instrument computer                                                  |
|      | Establish a method of transferring PDFs from the instrument computer to the analyst's computer |
|      | Demonstrate transferring PDF files from the instrument computer to the analyst's computer      |
|      | Acquire PDF editing software                                                                   |
|      | Demonstrate the merging of PDFs                                                                |
|      | Demonstrate the naming of PDFs                                                                 |
|      | Demonstrate how to make a correction to a PDF                                                  |
|      | Demonstrate uploading a PDF file into the BEAST Image Vault                                    |
|      | Demonstrate removing a PDF file from the BEAST Image Vault                                     |



# OSBI Drug Laboratory Training Manual

*Revision #20, Effective Date 06-15-2026*

Upon signing the approval, the trainee and trainer will review the above information and ensure the trainee has demonstrated knowledge and understanding of the above topics.

## **Approval**

Trainee \_\_\_\_\_ Date \_\_\_\_\_

Trainer/  
Supervisor \_\_\_\_\_ Date \_\_\_\_\_

Comments \_\_\_\_\_



# OSBI Drug Laboratory Training Manual

Revision #20, Effective Date 06-15-2026

## Drug Analysis by FTIR (DR-60)

### Goals

- To learn the theory of FTIR and how to use it as a confirmatory test in the analysis of submitted samples.
- To familiarize the analyst with how to maintain the FTIR instrument and ensure that it is working properly by using quality assurance and quality control methods.
- The analyst will learn how to interpret data from this type of analysis.

### Literature

| Date | Literature                                                                                                                                                                |
|------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|      | Protocol DR-60 (Drug Analysis by FTIR)                                                                                                                                    |
|      | Thermo Scientific. <b>FT-IR Glossary</b> . (PDF)                                                                                                                          |
|      | Thermo Scientific. <b>Introduction to Fourier Transform Infrared Spectroscopy</b> (PDF)                                                                                   |
|      | LCGC ChromAcademy. <b>Introduction to Infrared Spectroscopy</b> . (PDF)                                                                                                   |
|      | Perkin Elmer, <b>FT-IR Spectroscopy Attenuated Total Reflectance (ATR)</b> . (PDF)                                                                                        |
|      | Bell, S. <b>Spectroscopy</b> . <i>Forensic Chemistry</i> . pages 149-159                                                                                                  |
|      | Bell, S. <b>Infrared Spectroscopy</b> . <i>Forensic Chemistry</i> . pages 161-169                                                                                         |
|      | Hugel, J., Meyers, J.A. & Lankin, D.C. <b>Analysis of the Hallucinogens, Infrared (IR) Spectroscopy</b> . <i>Handbook of Forensic Drug Analysis</i> . 2005, pages 154-164 |
|      | <i>Clarke's Isolation and Identification of Drugs</i> . Vol. I, 3 <sup>rd</sup> Edition, pages 328-344                                                                    |

### Discussion

| Date                  | Tasks                                                                                                                                                                                                                               |
|-----------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| <b>Theory of FTIR</b> |                                                                                                                                                                                                                                     |
|                       | Theory and definition of FTIR                                                                                                                                                                                                       |
|                       | Define: <ul style="list-style-type: none"><li>• Wavelength</li><li>• Wavenumber</li><li>• Interferometer</li><li>• Constructive Interference (pertaining to FTIR)</li><li>• Destructive Interference (pertaining to FTIR)</li></ul> |



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|                 |                                                                                                                                                                                                                                                                                               |
|-----------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|                 | <p>Explain how the following pertain to FTIR</p> <ul style="list-style-type: none"> <li>• Gain</li> <li>• Resolution</li> <li>• Single beam spectrum</li> <li>• Sensitivity</li> </ul>                                                                                                        |
|                 | Describe the components of the FTIR and their function                                                                                                                                                                                                                                        |
|                 | The differences between transmission and reflectance modes                                                                                                                                                                                                                                    |
|                 | What wavelength range is analyzed using FTIR                                                                                                                                                                                                                                                  |
|                 | Explain evanescent wave as it pertains to FTIR ATR                                                                                                                                                                                                                                            |
|                 | How the interferometer functions                                                                                                                                                                                                                                                              |
|                 | Describe what a laser is and its function in FTIR, including its travel path within the FTIR                                                                                                                                                                                                  |
|                 | What is a background and why is it collected                                                                                                                                                                                                                                                  |
|                 | Describe what ATR is                                                                                                                                                                                                                                                                          |
|                 | <p>How the ATR functions including:</p> <ul style="list-style-type: none"> <li>• What the crystal is made of and why it doesn't interfere with analysis</li> <li>• Why CO<sub>2</sub> and H<sub>2</sub>O can still appear in a spectrum after background scans have been collected</li> </ul> |
|                 | Describe what the FTIR does to the sample during analysis                                                                                                                                                                                                                                     |
|                 | Why FTIR on tablets can be difficult                                                                                                                                                                                                                                                          |
| <b>Controls</b> |                                                                                                                                                                                                                                                                                               |
|                 | When spectra of a polystyrene standard will be obtained and what will it be compared to                                                                                                                                                                                                       |
|                 | The proper corrective procedure when the polystyrene standard fails                                                                                                                                                                                                                           |
|                 | When a background spectrum will be collected                                                                                                                                                                                                                                                  |
|                 | List ways contamination may be identified on the stage crystal and the tower arm                                                                                                                                                                                                              |
|                 | What steps are taken to ensure the ATR is free from contamination                                                                                                                                                                                                                             |
|                 | Describe the spectra of a "blank"                                                                                                                                                                                                                                                             |



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| Misc. |                                                                                                               |
|-------|---------------------------------------------------------------------------------------------------------------|
|       | Describe how to prepare a sample for FTIR analysis using the ATR                                              |
|       | Explain how humidity can affect the spectra and the quality of the match                                      |
|       | Explain how to remove moisture from a sample                                                                  |
|       | What must be done if analysis by FTIR does not indicate a controlled dangerous substance                      |
|       | Articulate why a second sample would be taken and what documentation is needed to be included in the casefile |
|       | Articulate reasons that FTIR data may be "rejected"                                                           |

## Tasks

| Date                           | Tasks                                                                                                                            |
|--------------------------------|----------------------------------------------------------------------------------------------------------------------------------|
| <b>Maintenance</b>             |                                                                                                                                  |
|                                | Demonstrate the analysis of a polystyrene standard which is to be analyzed using the "OSBI Macro," before casework is performed. |
|                                | Demonstrate proper cleaning of the ATR                                                                                           |
|                                | Demonstrate checking the humidity levels inside the instrument                                                                   |
| <b>Use of the Instrument</b>   |                                                                                                                                  |
|                                | Demonstrate preparation of a sample for analysis                                                                                 |
|                                | Demonstrate collection of background scans                                                                                       |
|                                | Demonstrate collection of scans using the ATR accessory                                                                          |
| <b>Spectral Interpretation</b> |                                                                                                                                  |
|                                | Review instrumental data and discuss what is and is not acceptable for casework analysis                                         |
|                                | Determine if CO <sub>2</sub> and/ or H <sub>2</sub> O are present in the scans                                                   |
|                                | Determine if another compound is present in the scans                                                                            |
|                                | Demonstrate proper report writing                                                                                                |



# OSBI Drug Laboratory Training Manual

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## **Approval**

Trainee \_\_\_\_\_ Date \_\_\_\_\_

Trainer/  
Supervisor \_\_\_\_\_ Date \_\_\_\_\_

Comments \_\_\_\_\_





# OSBI Drug Laboratory Training Manual

Revision #20, Effective Date 06-15-2026

## Requirements Prior to Drug Analysis Using FTIR

### Sample Analysis

| Date | Tasks                                                                                                                                            |
|------|--------------------------------------------------------------------------------------------------------------------------------------------------|
|      | Analyze at least 30 practice samples and record all results;<br>document using an Excel spreadsheet and archive in analyst's folder on QA server |

### Evaluation of Training

| Date | Tasks                                                                                                                             |
|------|-----------------------------------------------------------------------------------------------------------------------------------|
|      | Complete and review a competency test, with accurate results                                                                      |
|      | Complete a technical questions session with a minimum score of 80%, with<br>Technical Manager or Appointee<br>Average Score _____ |

### Approval

Trainee \_\_\_\_\_ Date \_\_\_\_\_

Trainer/  
Supervisor \_\_\_\_\_ Date \_\_\_\_\_

Comments \_\_\_\_\_



# OSBI Drug Laboratory Training Manual

Revision #20, Effective Date 06-15-2026

## Gas Chromatograph Mass Spectrometer Methods for Drug Analysis (DR-70)

### Goals

- To learn the theory of gas chromatography mass spectrometer and how to use it as a confirmatory test in the analysis of submitted samples.
- To familiarize the analyst with how to maintain the gas chromatograph mass spectrometer instrument and ensure that it is working properly by using quality assurance and quality control methods.
- The trainee will learn how to interpret data from this type of analysis.

**\*\*Prior to beginning, the trainee must complete GC training (DR-30)\*\***

### Literature

| Date | Literature                                                                                                                                                                                                                                                           |
|------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|      | Protocol DR-70 (Gas Chromatograph Mass Spectrometer Methods for Drug Analysis)                                                                                                                                                                                       |
|      | Basic Operation of the Mass Spectrometer (Appendix II of Training Manual)                                                                                                                                                                                            |
|      | Isomers (Appendix V of Training Manual)                                                                                                                                                                                                                              |
|      | Hugel, J., Meyers, J.A. & Lankin, D.C. <b>Analysis of the Hallucinogens, Mass Spectrometry.</b> <i>Handbook of Forensic Drug Analysis</i> . 2005, pages 176-185                                                                                                      |
|      | Pavia, D.L., Lampman, G.M., Kriz, G.S. <b>Mass Spectrometry.</b> <i>Intro to Spectroscopy</i> . 2001, pages 390-398 and 446-448                                                                                                                                      |
|      | Optional: Prall, J. D. & Cardone. <b>Gas Chromatography/Mass Spectrometric (GC/MS) Analysis of Drugs Using Spectral and Retention Index Matching.</b> DEA Central Lab, Dallas, TX and FAA Toxicology and Research Lab, OKC, OK.<br>(No date and reference available) |

### Discussion

| Date                               | Tasks                                                                                                                                                                                       |
|------------------------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| <b>Theory of Mass Spectrometry</b> |                                                                                                                                                                                             |
|                                    | The different methods and when to use for analysis, i.e., drug100, drug200, LSD, extnd100, low100, meth100, and/or any other methods being used for analysis                                |
|                                    | Split ratio and why it is used with the instrument                                                                                                                                          |
|                                    | Capillary columns used in the mass spectrometer <ul style="list-style-type: none"><li>• Nonpolar and polar liquid phases</li><li>• How does film thickness affect the performance</li></ul> |
|                                    | Splitting of the molecule and the function of the ion source                                                                                                                                |
|                                    | The quadrupole mass filter and how it functions                                                                                                                                             |
|                                    | The electron multiplier                                                                                                                                                                     |



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|                 |                                                                                                                                                                                                                     |
|-----------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|                 | Retention time and retention index, and how it applies to the GC/MS                                                                                                                                                 |
|                 | What is the hydrocarbon ladder and what role does it play in the retention index                                                                                                                                    |
|                 | How the retention index is calculated                                                                                                                                                                               |
|                 | The maximum allowed difference of the calculated Retention Index Difference                                                                                                                                         |
|                 | What carrier gas does the Drug Lab use                                                                                                                                                                              |
| <b>Controls</b> |                                                                                                                                                                                                                     |
|                 | What is the negative control and when should it be ran                                                                                                                                                              |
|                 | What is the positive control and when should it be ran                                                                                                                                                              |
|                 | When solvent blanks are to be run                                                                                                                                                                                   |
|                 | The proper corrective procedure if the retention index of the Cocaine standard exceeds plus or minus 2%                                                                                                             |
|                 | The proper corrective procedure if contamination occurs during the daily solvent blank run                                                                                                                          |
|                 | The proper corrective procedure if contamination occurs during a casework solvent blank run                                                                                                                         |
|                 | When the Tune Eval/Autotune are to be run                                                                                                                                                                           |
|                 | The proper corrective procedure if erroneous assignment of mass values to fragments in a sample, standard, or autotune                                                                                              |
|                 | The proper corrective procedure for the failure to produce a satisfactory cocaine spectrum for the cocaine standard                                                                                                 |
|                 | The proper corrective procedure if contamination peaks are found on the autotune (m/e 18, 44, etc.)                                                                                                                 |
|                 | When running the hydrocarbon ladder is required                                                                                                                                                                     |
|                 | When changing the liner and septum are required                                                                                                                                                                     |
|                 | When changing the gold seal is required and the difference between the GC and the GC/MS                                                                                                                             |
| <b>Misc.</b>    |                                                                                                                                                                                                                     |
|                 | What is the temperature programming and where can it be found                                                                                                                                                       |
|                 | The theory of a mass spectral library and where the libraries come from                                                                                                                                             |
|                 | When does a standard have to be run                                                                                                                                                                                 |
|                 | What information has to be retained with a new standard                                                                                                                                                             |
|                 | What information about the standard is listed on each mass spectra printout                                                                                                                                         |
|                 | Articulate reasons that data from the GC/MS may be "rejected"                                                                                                                                                       |
|                 | What happens to sample after analysis through instrument? <ul style="list-style-type: none"> <li>• Where does the waste from split vent go?</li> <li>• Where does sample go after leaving the mass spec?</li> </ul> |



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|  |                                                                                                                                                                                           |
|--|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|  | Define: <ul style="list-style-type: none"> <li>• Structural Isomers</li> <li>• Stereoisomers</li> <li>• Enantiomers/Optical Isomers</li> <li>• Diastereomers/Geometric Isomers</li> </ul> |
|--|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|

## Tasks

| Date                         | Tasks                                                                                                                                                                                                                                                                        |
|------------------------------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| <b>Maintenance</b>           |                                                                                                                                                                                                                                                                              |
|                              | Demonstrate how to record maintenance performed in the instrument maintenance log                                                                                                                                                                                            |
|                              | Demonstrate how to prepare and run the hydrocarbon ladder and ensure the appropriate retention times have been updated in the macro                                                                                                                                          |
|                              | Demonstrate changing liner and septum, resetting macro counts                                                                                                                                                                                                                |
|                              | Demonstrate changing gold seal, cleaning injection port and septa nut, and resetting macro count                                                                                                                                                                             |
|                              | Demonstrate how to autotune instrument, interpret the data and save the PDFs                                                                                                                                                                                                 |
|                              | Demonstrate how to vent the mass spec                                                                                                                                                                                                                                        |
|                              | Demonstrate how to disassemble and clean ion source, replace filaments, and reassemble                                                                                                                                                                                       |
|                              | Demonstrate how to pump down the mass spec                                                                                                                                                                                                                                   |
|                              | Demonstrate how to change the split vent filter and line                                                                                                                                                                                                                     |
|                              | Demonstrate how to change a column                                                                                                                                                                                                                                           |
| <b>Use of the Instrument</b> |                                                                                                                                                                                                                                                                              |
|                              | Demonstrate how to properly dilute or concentrate a sample for GC/MS analysis                                                                                                                                                                                                |
|                              | Demonstrate how to properly load a sample for analysis on the GC/MS, including entering information in the Sequence log                                                                                                                                                      |
|                              | Demonstrate the proper use of controls: <ul style="list-style-type: none"> <li>• Reagent Blanks</li> <li>• Cocaine Standards</li> </ul>                                                                                                                                      |
|                              | Articulate the requirements for Reagent Blanks and the Cocaine Standard <ul style="list-style-type: none"> <li>• What constitutes an acceptable Blank?</li> <li>• What constitutes an acceptable Cocaine Standard?</li> <li>• What methods are used and when/why?</li> </ul> |
|                              | Demonstrate the interpretation and comparison of the mass spectral data received from the instrument                                                                                                                                                                         |



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|  |                                                                                                                                                         |
|--|---------------------------------------------------------------------------------------------------------------------------------------------------------|
|  | Demonstrate how to perform a background subtraction and articulate when a background subtraction is needed.                                             |
|  | Demonstrate how to perform a manual scan. Articulate when a manual scan may be needed and what documentation is required if a manual scan is performed. |
|  | Demonstrate the proper reporting of results from the data received from the instrument                                                                  |

| Spectral Interpretation |                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                             |
|-------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|                         | <p>The trainee will review instrumental data and discuss with the trainer what is and is not acceptable for casework analysis</p> <ul style="list-style-type: none"><li>• Peaks past the molecular ion peak</li><li>• Background subtraction</li><li>• Complete spectra</li><li>• Extra/absent ions in a spectrum</li><li>• Calculate retention index difference</li></ul> <p><i>* See Training folder for examples of instrumental data that might need more work</i><br/><a href="#">\\VM-FSC-FILES\Common\DrugLab\1 Technical Manager\1 Training</a></p> |
|                         | What are possible sources of high background and how do you fix it                                                                                                                                                                                                                                                                                                                                                                                                                                                                                          |
|                         | Why would you experience broad, misshaped, or tailing peaks and how do you fix each one                                                                                                                                                                                                                                                                                                                                                                                                                                                                     |

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## Approval

Trainee \_\_\_\_\_ Date \_\_\_\_\_

Trainer/  
Supervisor \_\_\_\_\_ Date \_\_\_\_\_

Comments \_\_\_\_\_



# OSBI Drug Laboratory Training Manual

Revision #20, Effective Date 06-15-2026

## Administrative and Technical Reviewing of Casework

### Goals:

- To provide the trainee with knowledge and skills necessary to perform Administrative and Technical Reviews on another Analyst's casework

### Literature

| Date | Literature                  |
|------|-----------------------------|
|      | Review OSBI QP 31, Reviews  |
|      | Review DRQM-12 Case Reviews |

### Discussion

| Date | Tasks                                                                                                                                                             |
|------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|      | Discuss with Trainer how to perform a Technical Review of a Drug Case                                                                                             |
|      | Discuss how different prosecutorial charges affect the identification of certain compounds (i.e., pseudoephedrine vs. ephedrine, trafficking, distribution, etc.) |

### Tasks

| Date | Tasks                                                                    |
|------|--------------------------------------------------------------------------|
|      | Observe three Analysts perform Admin/Tech Reviews (5 cases each Analyst) |
|      | Name of Analyst 1:                                                       |
|      | Name of Analyst 2:                                                       |
|      | Name of Analyst 3:                                                       |

Upon signing the approval, the trainee and trainer will review the above information and ensure the trainee has demonstrated knowledge and understanding of the above topics.

### Approval

Trainee \_\_\_\_\_ Date \_\_\_\_\_

Trainer/  
Supervisor \_\_\_\_\_ Date \_\_\_\_\_

Comments \_\_\_\_\_



# OSBI Drug Laboratory Training Manual

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## Requirements Prior to Drug Analysis

### Sample Analysis

| Date | Tasks                                                                                                                                                                                                                                                                                                                                                                                                                                                     |
|------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|      | Perform the visual examination on approximately 25 green leafy samples that includes both negatives and positives, analyze using thin layer chromatography, and record all results; document using an Excel spreadsheet and archive in the analyst's folder on QA server.<br><i>All results must be accurate; if not then documentation of trainee &amp; trainer review must be completed with explanation of possible differences &amp; TM notified.</i> |
|      | Complete a competency for the calibrated pipettes. A satisfactory competency test will be within 10% of the expected value. The Technical Manager or trainer will prepare the competency and record the expected value.                                                                                                                                                                                                                                   |
|      | Analyze approximately 100 provided samples on the GC and GC/MS, to include positive and negative controlled dangerous substances and record all results; document using an Excel spreadsheet and archive in the analyst's folder on QA server.<br><i>All results must be accurate; if not then documentation of trainee &amp; trainer review must be completed with explanation of possible differences &amp; TM notified.</i>                            |
|      | Analyze approximately 70 practice drug cases, on the GC and GC/MS, and record all results; document using an Excel spreadsheet and archive in the analyst's folder on QA server.<br><i>All results must be accurate; if not then documentation of trainee &amp; trainer review must be completed with explanation of possible differences &amp; TM notified.</i>                                                                                          |
|      | Perform a training technical review on 50 cases.<br>These cases will be routed to the trainee in the BEAST using the route for training purposes code. The technical review will be documented on an Excel spreadsheet. The reviews done by the trainee will be compared to the official technical review by the TM or the Trainer.                                                                                                                       |

### Evaluation of Training

| Date | Tasks                                                                                                                          |
|------|--------------------------------------------------------------------------------------------------------------------------------|
|      | Complete and review a competency test, with accurate results                                                                   |
|      | Complete a technical questions session with a minimum score of 80%, with Technical Manager or Appointee<br>Average Score _____ |
|      | Complete a mock trial session, with approval from Technical Manager                                                            |



# OSBI Drug Laboratory Training Manual

*Revision #20, Effective Date 06-15-2026*

## Approval

Trainee \_\_\_\_\_

Date \_\_\_\_\_

Trainer/  
Supervisor \_\_\_\_\_

Date \_\_\_\_\_

Comments \_\_\_\_\_





# OSBI Drug Laboratory Training Manual

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## Liquid Nitrogen Safety

Liquid Nitrogen is the liquefied form of nitrogen gas. When nitrogen is in the gas phase, it is mostly inert gas that is colorless, odorless, and tasteless. In the liquid phase, nitrogen is very cold (-196 C or -320 F), which makes it ideal for keeping things cool. Because of its extremely cold temperature, any exposure to your skin can cause severe frostbite. On vaporization, liquid nitrogen expands by a factor of almost 700, so 1 liter of liquid nitrogen becomes 24.6 cubic feet of nitrogen gas. This can cause explosion of a sealed container, or it can displace oxygen in the room and cause suffocation without warning. Liquid nitrogen should always be stored in a vented container in a well-ventilated room. Oxygen may condense on the surface of liquid nitrogen causing it to be highly reactive with organic materials. This can cause ordinarily noncombustible materials to burn rapidly when it comes in contact with oxygen enriched liquid nitrogen. When handling liquid nitrogen, always wear thermal gloves and a protective face shield. Never dispose of liquid nitrogen by pouring it on the floor as it could displace enough oxygen to cause suffocation. Nitrogen gas is colorless and odorless, the cloud that forms when liquid nitrogen is poured on the floor is condensed water vapor from the air, not nitrogen gas.

## Goals

- To become knowledgeable of the hazards of liquid nitrogen
- To learn the safety precautions to utilize when handling liquid nitrogen
- To learn how to properly use/transfer liquid nitrogen

## Literature

| Date | Literature                                                                                         |
|------|----------------------------------------------------------------------------------------------------|
|      | Safety Data Sheet for liquid nitrogen                                                              |
|      | OSBI Policy 121.1 Appendix I, personal protective equipment required when handling liquid nitrogen |

## Discussion

| Date | Tasks                                                                         |
|------|-------------------------------------------------------------------------------|
|      | Why is liquid nitrogen used in the laboratory                                 |
|      | What are three hazards of liquid nitrogen                                     |
|      | What personal protective equipment must be used when handling liquid nitrogen |
|      | What is the proper type of container to use to transport liquid nitrogen      |
|      | What first aid is necessary if liquid nitrogen spills on skin or eyes         |
|      | How do you report a liquid nitrogen injury if it occurs                       |
|      | Why is there a safety release valve on the cryogenic cylinder                 |
|      | Why must you never use a tight-fitting cap on a dewar of liquid nitrogen      |



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## Tasks

| Date | Tasks                                                                                                                        |
|------|------------------------------------------------------------------------------------------------------------------------------|
|      | Trainer will demonstrate the proper technique for transferring liquid nitrogen from the cryogenic cylinder to a small dewar. |

## Evaluation of Training

|  |                                                      |
|--|------------------------------------------------------|
|  | Demonstrate how to properly transfer liquid nitrogen |
|--|------------------------------------------------------|

Upon signing the approval, the trainee and trainer will review the above information and ensure the trainee has demonstrated knowledge and understanding of the above topics.

### Approval

Trainee \_\_\_\_\_ Date \_\_\_\_\_

Trainer/  
Supervisor \_\_\_\_\_ Date \_\_\_\_\_

Comments \_\_\_\_\_



# OSBI Drug Laboratory Training Manual

Revision #20, Effective Date 06-15-2026

## Gas Chromatograph Infrared Detector Methods for Drug Analysis (DR-75)

### Goals

- To learn the theory of gas chromatography infrared detector and how to use it as a confirmatory test in the analysis of submitted samples.
- To familiarize the analyst with how to maintain the gas chromatograph infrared detector instrument and ensure that it is working properly by using quality assurance and quality control methods.
- The trainee will learn how to interpret data from this type of analysis.

***\*\*Prior to beginning, the trainee must complete GC (DR-30), FTIR training (DR-60), and Liquid Nitrogen Safety\*\****

### Literature

| Date | Literature                                                                     |
|------|--------------------------------------------------------------------------------|
|      | Protocol DR-75 (Gas Chromatograph Infrared Detector Methods for Drug Analysis) |
|      | IRD3 Operations Manual, Rev 1-3, ASIC                                          |
|      | Essential FTIR Operations Manual for GC IRD Users, Rev 0-3                     |
|      | IRD 3 Hardware & Schematic Overview                                            |

### Tasks

| Date | Tasks                                                                                                                                |
|------|--------------------------------------------------------------------------------------------------------------------------------------|
|      | <b>Maintenance</b>                                                                                                                   |
|      | Demonstrate how to fill the GCIRD with liquid nitrogen                                                                               |
|      | <b>Use of the Instrument</b>                                                                                                         |
|      | Demonstrate how to properly load a sample for analysis on the GCIRD, including entering information in the Sample Table              |
|      | Demonstrate the proper use of controls: <ul style="list-style-type: none"><li>• Solvent Blanks</li><li>• Cocaine Standards</li></ul> |
|      | Demonstrate the interpretation and comparison of data                                                                                |



# OSBI Drug Laboratory Training Manual

Revision #20, Effective Date 06-15-2026

## Requirements Prior to Drug Analysis using the GCIRD

### Sample Analysis

| Date | Tasks                                                                                                                                         |
|------|-----------------------------------------------------------------------------------------------------------------------------------------------|
|      | Analyze at least 30 practice samples and record all results; document using an Excel spreadsheet and archive in analyst's folder on QA server |

### Evaluation of Training

| Date | Tasks                                                        |
|------|--------------------------------------------------------------|
|      | Complete and review a competency test, with accurate results |

Upon signing the approval, the trainee and trainer will review the above information and ensure the trainee has demonstrated knowledge and understanding of the above topics.

### Approval

Trainee \_\_\_\_\_ Date \_\_\_\_\_

Trainer/  
Supervisor \_\_\_\_\_ Date \_\_\_\_\_

Comments \_\_\_\_\_



# OSBI Drug Laboratory Training Manual

Revision #20, Effective Date 06-15-2026

## Controlled Substances Technician

The Controlled Substances Technician's job is to assist the analysts and supervisors in the Controlled Substances Unit. This may include a variety of duties, including but not limited to: making reagents, checking scales & refrigerator temperatures, instrument maintenance and preparation of sampling apparatuses. Education and Experience Requirements: At a minimum, the Controlled Substances Technician is required to have graduated from high school or have an equivalent diploma. It is preferred the Technician has attended or is currently attending an accredited college or university, with preferred coursework in chemistry, forensic science, criminalistics, toxicology or a closely related natural science.

The Controlled Substances Technician will complete portions of the same sections of this training manual as a Criminalist in the Controlled Substances Unit. The trainee will date when each assignment is completed. The trainee and trainer will sign/date the Approvals and initial/date the Checklist when the required sections have been completed. Required sections will be listed in the Checklist.

The trainer will demonstrate to the Controlled Substances Technician some of the tasks that will be required duties; there may be no associated section in the training manual and no reading associated with those tasks. By signing off on the Checklist, the trainee has demonstrated their ability of completing the task to the trainer.

*A competency test must be completed, with anticipated results, prior to being released to create items that could be used for testing. The competency may include testing performed by an authorized analyst of the item(s) used for testing.*

The Technician may be authorized to perform some duties before all sections of the Checklist are completed.

| Section                                                        | Trainee<br>Initials | Trainer<br>Initials | Date |
|----------------------------------------------------------------|---------------------|---------------------|------|
| <b>Weights and Measures Utilized</b>                           |                     |                     |      |
| <i>Read DR-3</i>                                               |                     |                     |      |
| <i>Read DR-4 (DR-4 Attachments not required)</i>               |                     |                     |      |
| <i>Demonstrate how to properly use &amp; document weights:</i> |                     |                     |      |
| <i>Bench Scale</i>                                             |                     |                     |      |
| <i>Large Capacity Scale</i>                                    |                     |                     |      |
| <i>Analytical Scale</i>                                        |                     |                     |      |
| <i>Complete a 31-day Measurement Study on the three scales</i> |                     |                     |      |



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|                                                                                           |  |  |  |
|-------------------------------------------------------------------------------------------|--|--|--|
| <b>Thermometers</b>                                                                       |  |  |  |
| <i>All sections</i>                                                                       |  |  |  |
| <b>Extractions &amp; Handling</b>                                                         |  |  |  |
| <i>Literature</i>                                                                         |  |  |  |
| <i>Solution Preparation of 0.45 N Sodium Hydroxide</i>                                    |  |  |  |
| <i>Solution Preparation of Concentrated Sodium Bicarbonate</i>                            |  |  |  |
| <i>Solution Preparation of Concentrated Sodium Hydroxide</i>                              |  |  |  |
| <i>Determination of pH of solution</i>                                                    |  |  |  |
| <i>Demonstrate the procedure for basic extraction</i>                                     |  |  |  |
| <b>Color Tests</b>                                                                        |  |  |  |
| <i>Read DR-10</i>                                                                         |  |  |  |
| <i>Read DR-11</i>                                                                         |  |  |  |
| <i>Read DR-13</i>                                                                         |  |  |  |
| <i>Solution Preparation of Marquis Reagent</i>                                            |  |  |  |
| <i>Demonstrate the quality control verification and documentation for Marquis Reagent</i> |  |  |  |
| <i>Solution Preparation of Cobalt Reagent</i>                                             |  |  |  |
| <i>Demonstrate the quality control verification and documentation for Cobalt Reagent</i>  |  |  |  |
| <i>Demonstrate the quality control verification and documentation for Bates Test</i>      |  |  |  |
| <b>Gas Chromatography</b>                                                                 |  |  |  |
| <i>Read DR-30</i>                                                                         |  |  |  |
| <i>Maintenance Section of Training Manual</i>                                             |  |  |  |
| <b>Analysis of Mushrooms</b>                                                              |  |  |  |
| <i>Solution Preparation of Mushroom TLC Reagent</i>                                       |  |  |  |
| <i>Solution Preparation of Ehrlichs</i>                                                   |  |  |  |
| <i>Demonstrate the verification of the TLC Reagent and Ehrlichs</i>                       |  |  |  |
| <b>Drug Analysis by FTIR</b>                                                              |  |  |  |
| <i>Read DR-60</i>                                                                         |  |  |  |
| <i>Maintenance Section of Training Manual</i>                                             |  |  |  |
| <b>Gas Chromatograph Mass Spectrometer</b>                                                |  |  |  |
| <i>Read DR-70</i>                                                                         |  |  |  |
| <i>Maintenance Section of Training Manual</i>                                             |  |  |  |
| <b>Miscellaneous</b>                                                                      |  |  |  |
| <i>Recycling (shredded paper, cardboard, etc.)</i>                                        |  |  |  |
| <i>Refill the hydrogen generator bottles</i>                                              |  |  |  |
| <i>Check temperatures on the refrigerators</i>                                            |  |  |  |



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|                                                |  |  |  |
|------------------------------------------------|--|--|--|
| <i>Gases</i>                                   |  |  |  |
| <i>Checking Eyewashes and documenting</i>      |  |  |  |
| <i>Printing reports</i>                        |  |  |  |
| <i>Checking Safety Showers and documenting</i> |  |  |  |
| <i>Decontaminating phones and documenting</i>  |  |  |  |

Upon signing the approval, the trainee and trainer will review the above information and ensure the trainee has demonstrated knowledge and understanding of the above topics.

## Approval

Trainee \_\_\_\_\_

Date \_\_\_\_\_

Trainer/  
Supervisor \_\_\_\_\_

Date \_\_\_\_\_

Comments \_\_\_\_\_



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## Appendix I - Basic Operation of the Gas Chromatograph

In order to achieve maximum resolution between similar compounds on the gas chromatograph (GC), basic understanding of certain variables should be understood. Clear separation must be obtained for the mass spectrometer (MS) to distinguish between a mixture of molecular compounds and a single molecular compound. The capillary gas chromatograph has four methods to separate different molecules: pressure, length of column, type of column, and temperature. By making use of all of these variables together, most compounds will separate well.

Pressure regulation is controlled at the injection port and can be varied to meet certain applications by a feature called EPC (Electronic Pneumatic Control). There are three modes EPC can be used to separate types of molecules in a sample: split, splitless, and pulsed split.

*Split Mode:* During a split injection, a liquid sample is introduced into a heated inlet where it vaporizes rapidly. A small amount of the vapor enters the column while the major portion exits from the split / purge vent. Split injections are primarily used for high concentration samples when you can afford to lose most of the sample out of the split / purge vent. It is also used for samples that cannot be diluted.

*Splitless mode:* In this mode, the purge valve is closed during the injection and remains so while the sample is vaporized in the liner and transferred to the column. At a specified time after the injection, the purge valve opens to sweep any vapors remaining in the liner out the split vent. This avoids solvent tailing due to the large inlet volume and small column flow rate. Since the entire sample gets transferred onto the column, this mode is primarily used for samples of low concentration.

*Pulsed Split:* The pressure pulse modes increase inlet pressure just before the beginning of a run and returns it to the normal value after a specified amount of time. The pressure pulse sweeps the sample out of the inlet and into the column faster, reducing the chance for sample decomposition in the inlet. This is helpful for large molecular weight compounds that tend to linger around in the inlet and thus tend to get purged in other EPC modes. This method can also help to increase sensitivity by placing a larger amount of sample on the column while decreasing the possibility of samples staying in the injection port and causing contamination.

The resolving power of a column can be dependent on the length of the column. The longer the column, the greater the resolving power. Longer columns allow for more interaction from each molecule in the sample with the stationary phase. Resolution between similar compounds with small differences can be achieved by increasing the length. Columns can be purchased in many lengths, but the growing trend is toward smaller columns since larger columns are more expensive and greater resolution by other factors can compensate for less resolution from the column.





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Capillary columns are a glass tube with a silianized coating containing different functional groups. Hundreds of different types of columns exist, each with a different type of functional group that will separate different compounds of interest. Some stationary phases can select for nonpolar, polar, and even compounds with lone pairs of electrons. The silianized layer thickness has a direct effect on the retention and elution temperature for each sample compound. Thicker films retain compounds longer by maximizing the amount of time the compounds spend in the stationary phase. Thinner films allow compounds to pass through the column faster, most likely with less separating ability.

Temperature variations in the injection port and on the column are important for separation by capillary gas chromatography. The injection port is usually set at 290°C, which vaporizes samples upon introduction to the GC. The injection port is not part of the column; it's a chamber where the liquid sample can change into the gas phase before entering the column. The oven, that contains the column, is relatively cool at a starting temperature of 190°C less than that of the injection port. This is hot enough to allow the volatile solvent to remain as a gas, but cold enough to cause the less volatile compounds to return to a liquid state. Once the sample becomes a liquid, it deposits itself on the column and won't migrate until the oven temperature heats up to the compound's boiling temperature. When the sample is once again in its gas phase, it travels through the column's stationary phase and mobile phase, jumping between the two. Separation has occurred through the difference in boiling points and through the amount of time different molecules spend in the stationary phase while traveling through the column.

By utilizing these four variables, separation of compounds can be achieved in most cases. Observation of the retention time is valuable in determining the identity of a compound, by comparison with a known standard. Changing just one of these variables can influence the retention time of a compound. The chromatograph should not contain wide peaks, since two or more compounds could resemble one peak. Ideally, sample concentration should be enough to give a single, narrow peak.

## **References:**

1. Missouri State Highway Patrol Forensic Laboratory Chemistry Training Manual.
2. Hewlett-Packard GC/MS Product Software, August 1996.

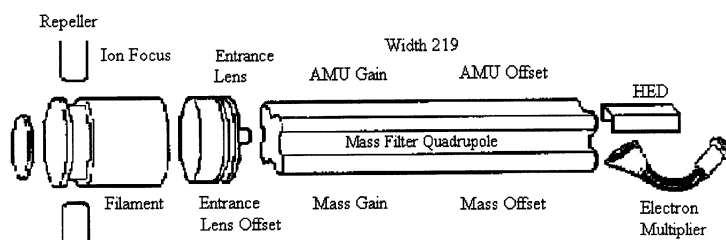


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## Appendix II - Basic Operation of the Mass Spectrometer

In order to interpret a tune report, basic understanding of the MS is necessary. The following is a brief explanation of how the Electron Impact (EI) ionization method works. The molecules come off of the GC column and are subjected to electron bombardment, which causes them to fragment and become charged. A repeller is used to direct the ions to the focusing lenses in the ion source and then to the quadrupole mass filter. The mass filter allows selected ion masses to reach the detector. It separates ions based on their masses allowing only ions of a specific mass to reach the detector at a given time. The quadrupole filters by applying to each pair of quadrupole rods, a combination of radio frequency (Rf) and direct current (DC) voltages. One rod pair receives Rf voltage 180 degrees out-of-phase with the other pair, while an equal but opposite DC potential is applied to each rod pair. Under these conditions, at any particular set



of Rf and DC voltage values, only ions of a specific mass to charge ( $m/z$ ) will traverse the length of the open space between the rods. All other ions are neutralized as they strike the surface of the rods. During a typical run, the MS scans

for masses ranging from 40 – 550 atomic mass units (amu). It scans for each mass unit in that range starting at the highest amu, working downward, throughout the duration of the run, with the exception of the solvent delay in which the MS is turned off. For instance, at the beginning of the scan, the mass filter selects only for masses of 550 amu, then it selects for masses of 549, and so on. This whole selection process takes place about three times a second. After each ion passes through the quadrupole, it is amplified by the electron multiplier, before reaching the detector. The detector counts the ions of each mass and plots the data on the mass spectrum (abundance vs. mass size). The quadrupole mass filter can select ions in two modes: Scan and SIM. Scan mode selects ions in the whole mass range specified, whereas SIM selects for specific mass units. The Scan mode has a lower sensitivity since most of the ions in a sample collide with the quadrupole rods. However, since samples are generally unknown, the filter mode utilized at the OSBI is the Scan mode to detect the entire spectrum of ions.

### Tuning

Tuning is the process for optimizing the performance of the Mass Selective Detector (MSD). The goal of tuning is to maximize sensitivity while maintaining acceptable resolution (the ability to distinguish between a mass and its isotope), ensuring accurate mass assignment, and providing the desired relative abundances across the spectrum. The Mass Spec (MS) uses Perfluorotributylamine (PFTBA) because its mass spectrum has ions in the low (69), medium (219), and high (502) mass range. The mass spectrum of PFTBA is shown on the bottom of the tune report. The instrument looks specifically for masses 69, 219, and 502 in the spectrum of PFTBA, and plots these values along a mass axis (the x-axis of the mass spectrum). The instrument



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aligns its internal mass axis to match the PFTBA mass axis. Resolution is determined by the ability of the instrument to distinguish two peaks, one mass unit apart. This resolution is displayed on the tune report by the peaks in the upper left-hand corner. The peak graph displays the mass of the peak, the abundance of that ion, and the peak width at 50% of the height (Pw50), as shown in the upper left portion of the graph. The x-axis of this graph is the mass assignment. By increasing the Pw50, the area at the base of the graph increases, and a larger range is allowed for the selected mass assignment (i.e., 69, 219, or 502). If made too large, the base area could encompass a range more than its peak range and label the isotope mass with the selected mass. This would achieve greater sensitivity, but the resolution would be very low since it could not distinguish the selected mass from its isotope. It is for that reason that the peak width must be between 0.4 and 0.6. While viewing the tune report, look for clear separation between the selected mass and its isotope.

On the upper right of the printout, there are numerous parameters displayed. These parameters are automatically assigned while using autotune to optimize the MSD performance. These values can be manually changed by using manual tune, but it is not recommended for normal use.

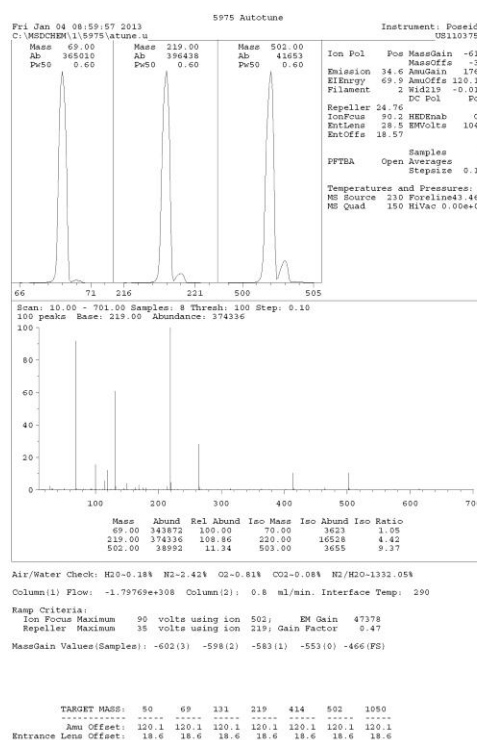
**Ion Pol:** This is the polarity of the field lens. A positive field pushes the ions out of the ion source. (5975)

**Emission:** The amount of current running through the filament. The higher the current the greater the electron bombardment but decreases a filament life. Too low of a current will result in less ionization and reduced sensitivity.

**ElEnerg:** The electron energy of the electron leaving the filament. (5975)

**Filament:** The MS contains two filaments in case one burns out.

**Repeller:** Sets the voltage of the repeller (part of the ion source). The repeller is a positive potential that repels the ions, pushing them out of the source. If the repeller is set too low, too few ions will leave the source, resulting in poor sensitivity and poor high mass response. If it is set too high, too many ions at too high a velocity will leave the source. This results in poor mass filtering and poor low mass resolution.





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- IonFcus:** Sets the voltage of the ion focus lens (part of the ion source). Ion focus affects ion abundance. Generally, the offset is ramped during the tuning to find the ion focus offset that results in the best ion abundance.
- EntLens:** Refers to the entrance lens gain, a value used to determine a mass dependent voltage that is applied to the entrance lens. The entrance lens is the final lens through which ions pass before they enter the mass filter quadrupole. Typically, during tuning, the entrance lens voltage is ramped to find the setting that provides the maximum abundance.
- EntOffs:** This is a constant voltage that is applied to the entrance lens. Increase the offset to increase abundance of ions at low masses without substantially decreasing the abundance of ions at high masses.
- PFTBA:** The status of the valve containing the PFTBA. This valve will open and close automatically for tuning.
- MassGain:** Sets the value of the mass axis gain, which is a multiplicative factor used in the equation to calibrate the mass axis. Mass gain adjusts the reported value of a given mass to the correct number. The mass that appears in a report has had a linear correction applied to it. This may be thought of as a calibration curve where the uncorrected mass is plotted along the x-axis and the reported mass is plotted along the y-axis. The calibration curve is a straight line with a slope that is proportional to the mass gain. Mass gain has a greater effect on mass assignments at the high end of the mass scale than at the low end.
- MassOffs:** This is an additive factor used in the equation to calibrate the mass axis.
- AMUGain:** Atomic mass unit gain affects the width of the mass peak by adjusting the ratio of DC voltage to RF voltage on the mass filter. A higher value gives narrower peaks, but affects peaks at high masses more than those at low masses.
- AMUOffs:** This affects the width of the mass peaks by adjusting the ratio of DC voltage of the mass filter quadrupole. A higher value gives narrower peaks at all masses.
- Wid219:** Affects the width of the mass peak at 219 amu. The value entered for this parameter is approximately the value of the correction applied at mass 219. For instance, if a peak width adjustment has been performed and the values are: Mass 69 Pw0.60, Mass 219 Pw0.63, Mass 502 Pw0.60, then entering a value of –0.03 for the Wid219 parameter, followed by a peak width adjustment, should result in the peak widths of all masses being set very close to 0.60 amu.



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- DC Pol:** Sets the polarity of the direct current applied to the quadrupole mass filter. This parameter is set at the optimum polarity at the factory and should not be changed for normal use.
- HEDEnab:** The High Energy Dynode sets the voltage to focus the ions into the detector, which is located off-axis, hidden from photons and electrons coming from the source. The optimal HED voltage depends on the electron multiplier setting. Thus, the electron multiplier voltage is usually set first. Then the HED voltage is ramped to determine the setting that provides the greatest abundance. The older instruments assigned a value to this parameter, which use to be called X-ray lens, however the HP 5975 MSD does not have an X-ray lens and just indicates "on" or "off". (5975)
- EMVolts:** The electron multiplier increases the abundance of all ions in the scan range going to the detector.
- Samples:** The log2 of the number of samples to be taken and averaged at each mass during a scan.
- Averages:** The number of profile scans to be averaged for each scan reported.
- Stepsize:** The mass axis increment used for a profile Scan. The larger the number, the faster scans are taken, at a cost of resolution.

## Temperatures and Pressures:

### **MS Source/**

**MSQuad:** Displays temperature settings for the Source and Quadrupole. (5975)

**Foreline:** The pressure between the rough pump and the diffusion pump. This area will either state the pressure of the foreline if the MSD uses a diffusion pump or the speed of the turbo pump. (5975)

**HiVac:** Displays the high vacuum pressure. (5975)



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On the Display of the mass spectrum of PFTBA, other parameters are listed:

**Scan:** 10.00-700.00 amu is the scan range during the tune. Typically, when drug samples are scanned, this parameter is approximately 40-550 amu.

**Samples:** The log2 of the number of samples to be taken and averaged at each mass during a scan. If the number is 8, log2 would be 256 scans.

**Threshold:** Abundances below this value will be ignored for scanning. This determines what signal will be accepted as peaks.

**Base:** Shows the base peak in the sample.

**Abundance:** Abundance of the base peak.

## Tune Evaluation (Tune Eval)

Tune evaluation is a way of verifying the performance of the MSD. First, it will evaluate the most current autotune (ATUNE.U) parameters and when the evaluation is complete, a system verification report is printed.

If all parameters of the autotune are within the predetermined limits, set by Agilent, they will be listed as "OK." If all parameters pass, the instrument can be used for casework. If any of the parameters fail, the reason for failure must be determined and corrected. The Autotune and Tune Evaluation must be run again and pass all parameters before casework can be analyzed on the instrument.

```
System Verification - Tune (Detector Optimization) Portion

Instrument Name      : Poseidon
DC Polarity         : Positive
Filament            : 2
BasePeak should be 69 or 219
Position of mass 69      69.00   OK
Position of mass 219     219.00   OK
Position of mass 502     502.00   OK
Position of isotope mass 70  70.00   OK
Position of isotope mass 220 220.00   OK
Position of isotope mass 503 503.00   OK
Ratio of mass 70 to mass 69(0.5 - 1.6%) 1.09   OK
Ratio of mass 220 to mass 219(3.2 - 5.4%) 4.38   OK
Ratio of mass 503 to mass 502(7.9 - 12.3%) 10.03   OK
Ratio of 219 to 69 should be > 40% and is 108.66   OK
Ratio of 502 to 69 should be > 2.4% and is 11.43   OK

Mass 69 Precursor (<= 3%) 0.17   OK
Mass 219 Precursor (<= 6%) 0.78   OK
Mass 502 Precursor (<= 12%) 1.35   OK

Testing for a leak in the system
Ratio of 18 to 69 (<20%) 0.16   OK
Ratio of 28 to 69 (<10%) 2.39   OK

Electron Multiplier Voltage 1047   OK

Tune portion of System Verification passed.
```

System Verification for Poseidon Fri Jan 04 09:03:17 2013 Page 1

## References:

1. Missouri State Highway Patrol Forensic Laboratory Chemistry Section Training Manual.
2. Hewlett-Packard GC/MS Product Software, August 1996.
3. [www.agilent.com](http://www.agilent.com)



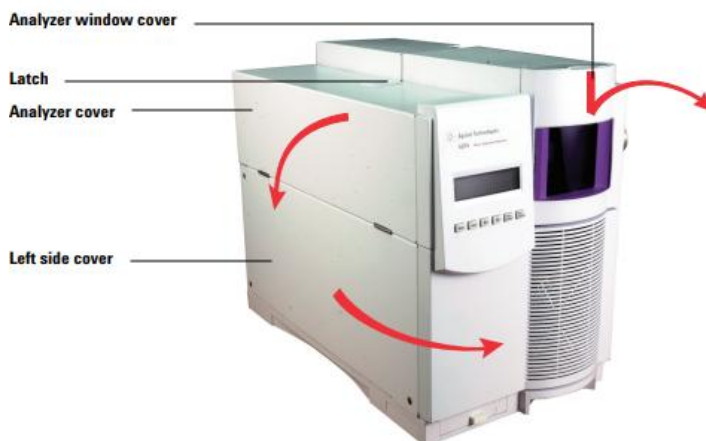
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## Appendix III – Helpful Tips for The Mass Spectrometer

### Pump down/Filament/Vent

1. In the Instrument Control view, select **Instrument**, select **MS Vacuum Control** to display the Vacuum Control dialog box or select **View**, select **Tune Vacuum Control**, select **Vacuum** select **MS Vacuum control**
2. Click **Vent**
  - 2.1. Follow the instructions presented on the screen, and wait until the vent cycle is completed.
  - 2.2. Let the rough pump cool off approximately 45 minutes if needed unplug the pump
  - 2.3. Close the software and turn off the MS portion on the instrument
3. Turn the vent valve counterclockwise only 3/4 turns or until you hear the hissing sound of air flowing into the analyzer chamber



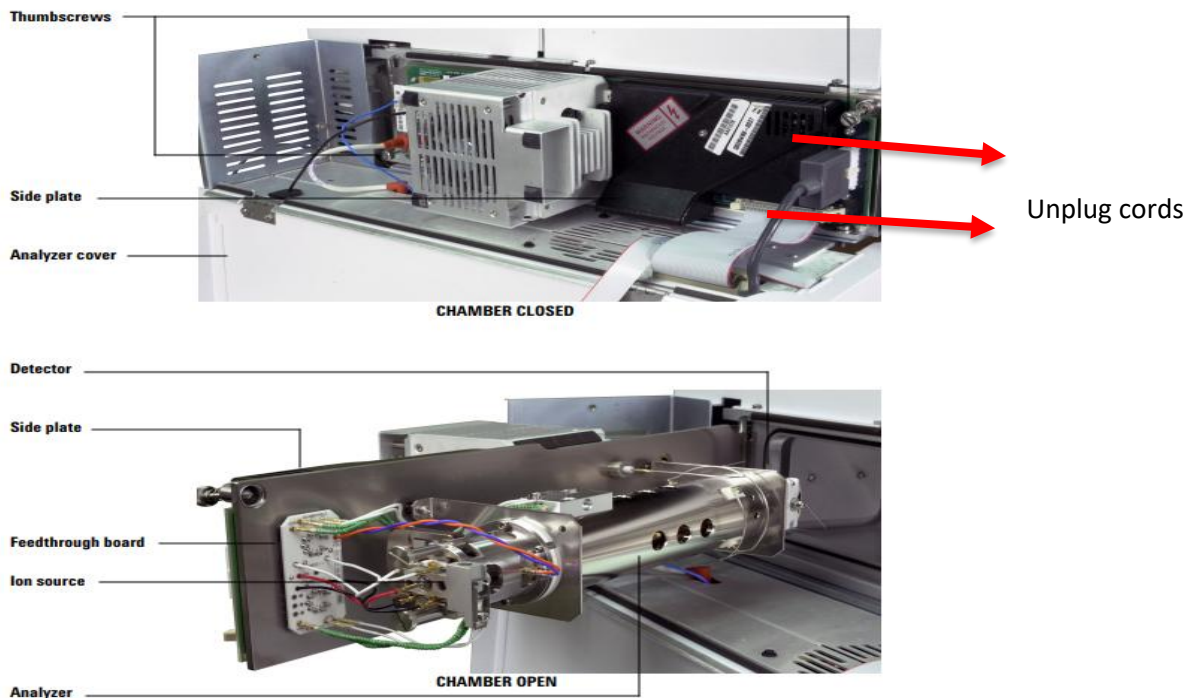
Vent valve knob



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4. Disconnect the side board control cable and the source power cable from the side board. Loosen the side plate thumbscrews if they are fastened and gently swing the side plate out



5. Remove the source from the analyzer
6. Disassemble the source and collect the parts to be cleaned. Clean using microgrit /sandpaper. Sonicate metal pieces using methanol approximately 15 minutes. Ensure microgrit is cleaned off before reassembling the source.

**Side note:** Ensure the new filaments are placed the correct way!



7. Reinstall the source into the analyzer and close the side plate
8. Reconnect the side board control cable and source power cable to the side board
9. Plug the rough pump back in





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10. Make sure the vent valve is closed turn knob clockwise
11. Press the Power on button on the front of the MS while pressing on the side plate to ensure a good seal
12. Start the Masshunter Data Analysis program. In the Instrument Control view, select **Instrument**, select **MS Vacuum Control** to display the Vacuum Control dialog box or select **View**, select **Tune Vacuum Control**, select **Vacuum** select **MS Vacuum control**
13. Select Pump Down
14. Allow ample time for instrument to pump down

## Switch Filaments

1. From the Instrument Control menu, select **View** then select **Tune & Vacuum Control**
2. Select **Parameters** next select **Manual Tune**
3. Enter the filament number
4. Click Done and save the atune file or select **Save Tune Parameters** from the **File** menu

## Manual Tune for air leak

1. From the Instrument Control menu, select **View** then select **Tune & Vacuum Control**
2. Select **Parameters** next select **Manual Tune**
3. Select the **Scan** tab of the **Manual Tune** dialog box and set the Scan Mass Range to Low  $m/z$  10 to High  $m/z$  100 also check that the **PFTBA** is set to **Closed** in the **Parameters** section.
4. Click **Scan**, look at the window labeled scan
5. Do not save atune

## Search Drug library

1. From the Instrument Control menu select **View**, select **MSD Chemstation Data Analysis**
2. Select **View** next select **Parametric Retrieval**
3. Enter in the library you would like to search default: *c:\Database\CrDM.l* **Note:** Other libraries are available to be searched
4. Choose your search option in the side panel

[5975 Series MSD Operation Manual for MassHunter \(agilent.com\)](http://www.agilent.com)



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## Appendix IV - Mass Peaks of Common Contaminants

| Mass(es)                                   | Compound General Classification | Potential Source                                                        |
|--------------------------------------------|---------------------------------|-------------------------------------------------------------------------|
| 18, 28, 32, 40, 44                         | Air                             | H <sub>2</sub> O, N <sub>2</sub> , O <sub>2</sub> , Ar, CO <sub>2</sub> |
| 18                                         | Cleaning Solvents               | Water                                                                   |
| 31                                         |                                 | Methanol                                                                |
| 47, 83, 85                                 |                                 | Chloroform                                                              |
| 77                                         |                                 | Benzene or Xylenes                                                      |
| 91,92                                      |                                 | Toluene                                                                 |
| 105,106                                    |                                 | Xylenes                                                                 |
| 43, 58                                     |                                 | Acetone                                                                 |
| 85                                         |                                 | Freons                                                                  |
| 73, 147, 207, 222, 281, 295, 341, 355, 429 | Dimethylpolysiloxane            | Septum or Column bleed                                                  |
| 41, 43, 55, 57, 71, 85, 99                 | Hydrocarbons                    | Fingerprints or pump oil                                                |
| 149                                        | Phthalates                      | Plasticizers in tubing, vials, caps, samples                            |



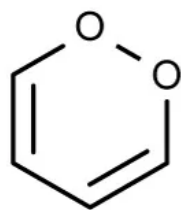
# OSBI Drug Laboratory Training Manual

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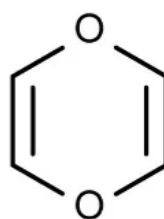
## Appendix V – Isomers

**Structural isomers:** a form of isomers in which molecules with the same Molecular Formula have different bonding patterns and atomic organization.

- Structural isomers share the same chemical formulas, but their atoms are arranged differently.
- Examples include ortho- para- meta-



*ortho-dioxin*

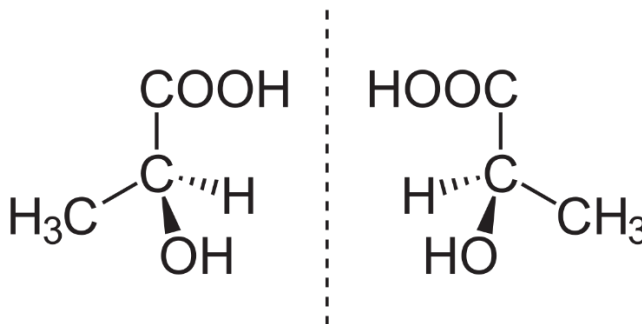


*para-dioxin*

**Stereoisomers:** a form of isomers in which molecules with the same Molecular Formula have molecular bonds which are always in the same order and only differ in spatial arrangement.

- Isomers that have the same number of the same kinds of atoms but differ in the orientation of their atoms in space. These isomers differ in chemical and physical properties.
- There are two types of stereoisomers: enantiomers and diastereomers.

**Enantiomers or Optical isomers:** 2 isomer molecules that are chiral, and are mirror images of each other; a pair of chemical compounds whose molecular structures have a nonsuperimposable mirror-image relationship to each other

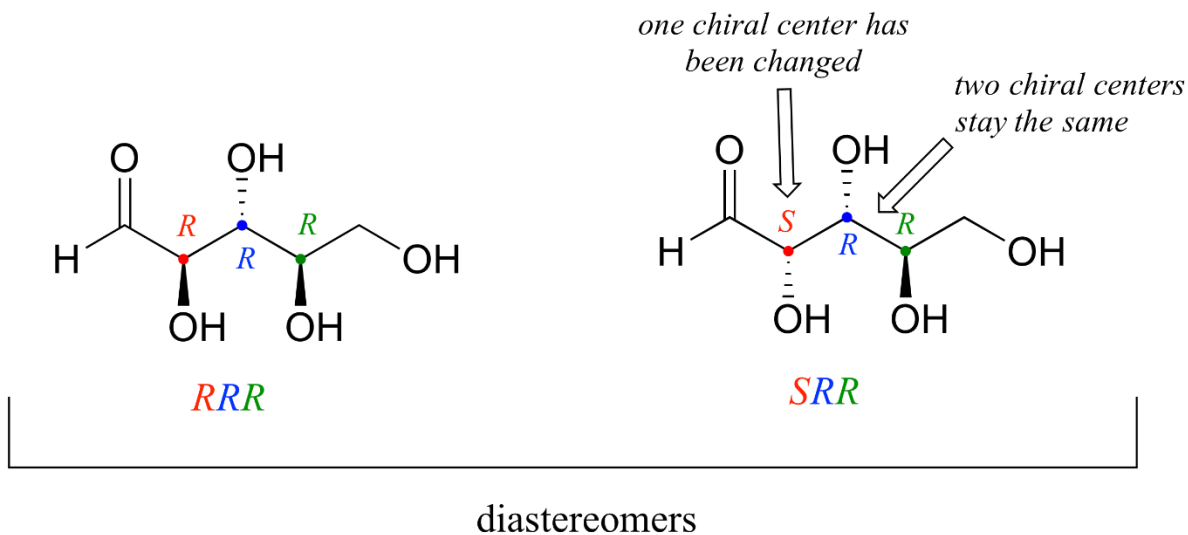




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**Diastereomers or Geometric isomers:** 2 stereoisomers that are not mirror images; a stereoisomer of a compound having two or more chiral centers that is not a mirror image of another stereoisomer of the same compound





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## Appendix VI – Casework Guidelines

These are just guidelines. Each case is different and might require more or less analysis. If you have questions about what to analyze or how to analyze, contact the Technical Manager (TM) or your Supervisor.

### Residues and Paraphernalia:

- If there is a weighable substance present you do not need to analyze the residue or paraphernalia.
- If the residue or paraphernalia is the only item linked to a specific suspect, it will need to be analyzed.
- If the case only contains residues, and it is not specified which suspect they belong to, only one of each substance type needs to be analyzed.

Example: Three bags of crystal residue, two bags of black tar residue, and two bags of green leafy residue.

Analyze: one bag with a crystal residue, one bag of black tar residue, and one bag of green leafy residue.

### Number of Items:

This needs to be looked at on a case-by-case basis.

- For trafficking case, analyze the number of items necessary to reach the trafficking level.
  - If performing a gross weight, weigh over the trafficking level to account for the weight of the packaging.
- For intent to distribute, analyze half or no more than 10 of the submitted items.
- For cases that do not involve trafficking or distribution:
  - Is there a CDS present?
  - Did you analyze each of the different types of substances?
- If the case contains weighable amounts of several different types of substances, one of each substance type needs to be analyzed.



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## **THC and mushroom products/food:**

This needs to be looked at on a case-by-case basis.

- If marijuana is present, you do not need to analyze the THC products/food.
- If mushrooms are present and test positive for psilocyn and/or psilocybin then you do not need to analyze the mushroom products/food.
- If there is not plant material present (marijuana or mushrooms) then test one to five items following the appropriate protocol.
  - If you have questions about the number of items to analyze talk to the TM or your Supervisor.
- If mushrooms or marijuana are not present but another CDS is then test one of the THC/mushroom products/foods.



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## Trafficking Marijuana Cases and the Matrix

There are two different ways that are acceptable:

1. You can fill out all the information for every item.

|                                                                  |  |                              |  |                                  |  |
|------------------------------------------------------------------|--|------------------------------|--|----------------------------------|--|
| Entry 1                                                          |  |                              |  |                                  |  |
| Item Number                                                      |  | Bundles Sampled              |  |                                  |  |
| 1                                                                |  |                              |  |                                  |  |
| Description                                                      |  |                              |  |                                  |  |
| One plastic bag containing a green leafy substance               |  |                              |  |                                  |  |
| Weight Type                                                      |  | Measurement                  |  | Units                            |  |
| net weight                                                       |  | 4.05                         |  | grams                            |  |
| Form                                                             |  | Residue Collection           |  |                                  |  |
| G (gls)                                                          |  |                              |  |                                  |  |
| UOM                                                              |  | Upper Weight for Trafficking |  | Upper Weight Unit                |  |
|                                                                  |  |                              |  |                                  |  |
| Lower Weight for Trafficking                                     |  | Lower Weight Unit            |  |                                  |  |
|                                                                  |  |                              |  |                                  |  |
| Marquis                                                          |  | Marquis Control              |  | Cobalt Thiocy / Bates            |  |
|                                                                  |  |                              |  |                                  |  |
| Cobalt Thiocy/Bates Control                                      |  |                              |  |                                  |  |
|                                                                  |  |                              |  |                                  |  |
| Literary Reference                                               |  | Literary Ref Result          |  | Amount                           |  |
|                                                                  |  |                              |  |                                  |  |
| Extraction                                                       |  | Instrumentation              |  |                                  |  |
| Methanol                                                         |  | GC / GCMS                    |  |                                  |  |
| Extraction 2                                                     |  | Instrumentation 2            |  |                                  |  |
|                                                                  |  |                              |  |                                  |  |
| Duquenois-Levine                                                 |  | TLC                          |  | Marihuana Visual Examination     |  |
|                                                                  |  | Positive: THC                |  | positive for cystolithic hairs   |  |
| Duquenois-Levine Control                                         |  | TLC Control                  |  | TLC Reagent/Visualization Agent  |  |
|                                                                  |  | Pass                         |  | 3:1 Hexanes:Acetone/Fast Blue BB |  |
| Date of Analysis                                                 |  | # of TLC Plates Used         |  | # of TLC Plates Passing QC       |  |
| 1/13/2026                                                        |  | 1                            |  | 1                                |  |
| pH                                                               |  |                              |  |                                  |  |
|                                                                  |  |                              |  |                                  |  |
| Final Result                                                     |  |                              |  |                                  |  |
| Cannabis (Marijuana containing Tetrahydrocannabinol), Schedule I |  |                              |  |                                  |  |
| Final Result 2                                                   |  |                              |  |                                  |  |
|                                                                  |  |                              |  |                                  |  |
| Final Result 3                                                   |  |                              |  |                                  |  |
|                                                                  |  |                              |  |                                  |  |
| Unconfirmed Compound #1                                          |  | Unconfirmed Compound #2      |  |                                  |  |
|                                                                  |  |                              |  |                                  |  |
| Notes                                                            |  |                              |  |                                  |  |
| EE                                                               |  |                              |  |                                  |  |

2. You can fill out all the information on the first item and:
  - a. Make a note on the first item for the extraction, instrumentation, TLC results, and visual exam for all items analyzed. (See Item 1 below)  
Ex. 1A – 1Z: All were extracted in MeOH and analyzed on the GC/GCMS. All were positive for delta 9-THC and Cystolithic hairs.



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- b. The matrix for every item should include the description, weight, form, and final results. (See Item 2 below)

|                                                                                                                                              |                              |                                  |                                  |                    |  |
|----------------------------------------------------------------------------------------------------------------------------------------------|------------------------------|----------------------------------|----------------------------------|--------------------|--|
| Entry 1                                                                                                                                      |                              |                                  |                                  |                    |  |
| Item Number                                                                                                                                  | Bundles Sampled              |                                  |                                  |                    |  |
| 1                                                                                                                                            |                              |                                  |                                  |                    |  |
| Description                                                                                                                                  |                              |                                  |                                  |                    |  |
| One plastic bag containing a green leafy substance                                                                                           |                              |                                  |                                  |                    |  |
| Weight Type                                                                                                                                  | Measurement                  | Units                            | Form                             | Residue Collection |  |
| net weight                                                                                                                                   | 4.05                         | grams                            | G (gls)                          |                    |  |
| UOM                                                                                                                                          | Upper Weight for Trafficking | Upper Weight Unit                | Lower Weight for Trafficking     | Lower Weight Unit  |  |
|                                                                                                                                              |                              |                                  |                                  |                    |  |
| Marquis                                                                                                                                      | Marquis Control              | Cobalt Thiocyanate / Bates       | Cobalt Thiocyanate/Bates Control |                    |  |
|                                                                                                                                              |                              |                                  |                                  |                    |  |
| Literary Reference                                                                                                                           | Literary Ref Result          | Amount                           |                                  |                    |  |
|                                                                                                                                              |                              |                                  |                                  |                    |  |
| Extraction                                                                                                                                   | Instrumentation              |                                  |                                  |                    |  |
| Methanol                                                                                                                                     | GC / GCMS                    |                                  |                                  |                    |  |
| Extraction 2                                                                                                                                 | Instrumentation 2            |                                  |                                  |                    |  |
|                                                                                                                                              |                              |                                  |                                  |                    |  |
| Duquenois-Levine                                                                                                                             | TLC                          | Marihuana Visual Examination     |                                  |                    |  |
|                                                                                                                                              | Positive: THC                | positive for cystolithic hairs   |                                  |                    |  |
| Duquenois-Levine Control                                                                                                                     | TLC Control                  | TLC Reagent/Visualization Agent  |                                  |                    |  |
|                                                                                                                                              | Pass                         | 3:1 Hexanes:Acetone/Fast Blue BB |                                  |                    |  |
| Date of Analysis                                                                                                                             | # of TLC Plates Used         | # of TLC Plates Passing QC       | pH                               |                    |  |
| 1/13/2026                                                                                                                                    | 1                            | 1                                |                                  |                    |  |
| Final Result                                                                                                                                 |                              |                                  |                                  |                    |  |
| Cannabis (Marijuana containing Tetrahydrocannabinol), Schedule I                                                                             |                              |                                  |                                  |                    |  |
| Final Result 2                                                                                                                               |                              |                                  |                                  |                    |  |
|                                                                                                                                              |                              |                                  |                                  |                    |  |
| Final Result 3                                                                                                                               |                              |                                  |                                  |                    |  |
|                                                                                                                                              |                              |                                  |                                  |                    |  |
| Unconfirmed Compound #1                                                                                                                      |                              | Unconfirmed Compound #2          |                                  |                    |  |
|                                                                                                                                              |                              |                                  |                                  |                    |  |
| Notes                                                                                                                                        |                              |                                  |                                  |                    |  |
| EE; Items 1 through 5: All items were extracted in MeOH and analyzed on the GC/GCMS. All were positive for delta 9-THC and cystolithic hairs |                              |                                  |                                  |                    |  |





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|                                                                  |                              |                                 |                              |                    |
|------------------------------------------------------------------|------------------------------|---------------------------------|------------------------------|--------------------|
| Entry 1                                                          |                              |                                 |                              |                    |
| Item Number                                                      | Bundles Sampled              |                                 |                              |                    |
| 2                                                                |                              |                                 |                              |                    |
| Description                                                      |                              |                                 |                              |                    |
| One plastic bag containing a green leafy substance               |                              |                                 |                              |                    |
| Weight Type                                                      | Measurement                  | Units                           | Form                         | Residue Collection |
| net weight                                                       | 6.84                         | grams                           | G (gls)                      |                    |
| UOM                                                              | Upper Weight for Trafficking | Upper Weight Unit               | Lower Weight for Trafficking | Lower Weight Unit  |
|                                                                  |                              |                                 |                              |                    |
| Marquis                                                          | Marquis Control              | Cobalt Thiocy / Bates           | Cobalt Thiocy/Bates Control  |                    |
|                                                                  |                              |                                 |                              |                    |
| Literary Reference                                               | Literary Ref Result          | Amount                          |                              |                    |
|                                                                  |                              |                                 |                              |                    |
| Extraction                                                       | Instrumentation              |                                 |                              |                    |
|                                                                  |                              |                                 |                              |                    |
| Extraction 2                                                     | Instrumentation 2            |                                 |                              |                    |
|                                                                  |                              |                                 |                              |                    |
| Duquenois-Levine                                                 | TLC                          | Marihuana Visual Examination    |                              |                    |
|                                                                  |                              |                                 |                              |                    |
| Duquenois-Levine Control                                         | TLC Control                  | TLC Reagent/Visualization Agent |                              |                    |
|                                                                  |                              |                                 |                              |                    |
| Date of Analysis                                                 | # of TLC Plates Used         | # of TLC Plates Passing QC      | pH                           |                    |
|                                                                  |                              |                                 |                              |                    |
| Final Result                                                     |                              |                                 |                              |                    |
| Cannabis (Marijuana containing Tetrahydrocannabinol), Schedule I |                              |                                 |                              |                    |
| Final Result 2                                                   |                              |                                 |                              |                    |
|                                                                  |                              |                                 |                              |                    |
| Final Result 3                                                   |                              |                                 |                              |                    |
|                                                                  |                              |                                 |                              |                    |
| Unconfirmed Compound #1                                          |                              | Unconfirmed Compound #2         |                              |                    |
|                                                                  |                              |                                 |                              |                    |
| Notes                                                            |                              |                                 |                              |                    |
| EE                                                               |                              |                                 |                              |                    |



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## Mock Trial Evaluation Form

Analyst \_\_\_\_\_

Score \_\_\_\_\_

Reviewer \_\_\_\_\_

Date \_\_\_\_\_

*Please rate the trainee's performance during the Mock Trial:*

|                                                           | Excellent<br>(3)         | Good<br>(2)              | Fair<br>(1)              | Poor<br>(0)              |
|-----------------------------------------------------------|--------------------------|--------------------------|--------------------------|--------------------------|
| Courtroom demeanor and appearance                         | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| Ability to convey information in an understandable manner | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| Poise and professionalism during direct examination       | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| Poise and professionalism during cross examination        | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| Use of court exhibits/visual aids (if applicable)         | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| Testimony based upon scientific principles                | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| Exhibition of knowledge of OSBI testing procedures        | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| Explanation of results                                    | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |

Remarks/Comments/Suggestions/Explanation for Poor Ratings: \_\_\_\_\_

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This section is intended to list articles/books/journals/etc. that are required reading, additionally training courses can be listed as well.

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## Approval

Technical  
Manager

Michelle Carter

Date 6-9-2026

Quality  
Manager

Dan E. Roca

Date 06/09/26

Criminalistics  
Division  
Director

Hampton

Date 06/09/2026

Comments

\_\_\_\_\_



# OSBI Drug Laboratory Training Manual

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## History

| Revision | Issue Date | History                                                                                                                                                                                                                                                                                                                                                                                       |
|----------|------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| 20       |            | <p><b>Extractions &amp; Handling:</b> Added "Review the extraction references in the Training folder." These were the references removed from DR-110.</p> <p><b>Appendix III:</b> Update step 1 for Switch Filaments and Manual Tune for air leak.</p> <p><b>Appendix VI:</b> Incorporated from deviation. Added examples for how to fill out the matrix for trafficking marijuana cases.</p> |