PRE-SAMPLING STRATEGY SESSION

♦ LABEL ALL PRIMARY (sterile) and SECONDARY (non-sterile) SAMPLE CONTAINERS:

1) In the cold zone label each primary and secondary container by writing:
   a. Type of sample- i.e. LETTER, DRY SWAB, BULK POWDER, LIQUID, etc.
   b. Date of collection, sample ID#, and sampler’s initials

♦ SAMPLING MATERIALS (also see Sampling Tools and Containers sheet):

1) Gloves
   a. Assistant Sampler: 2 pairs of non-powdered latex, nitrile or vinyl examination gloves
   b. Sampler: 1 base pair and 1 pair for each sample site

2) Plastic sheet or clean drop cloth for staging area. Stage away from sample area

3) 10% bleach solution in spray bottle

4) Red biohazard bag for carry sampling tools downrange and discarding materials

5) PRIMARY SOURCE SAMPLING ONLY (letter, small opened package, etc):
   a. Sterile primary container (unopened baggie; see above labeling instructions)
   b. Non-sterile secondary container (baggie; see above labeling instructions)

6) BULK POWDER SAMPLING ONLY (visible powder on a smooth, non-porous surface):
   a. Sterile laminated sampling card
   b. Sterile primary container (wide mouth, screw top plastic cup; see above labeling instructions)
   c. Sterile macrofoam swab
   d. Sterile primary container (plastic swab tube, screw top; remove plastic to label)
   e. Non-sterile self-sealing bags (see above labeling instructions)

7) VACUUM SAMPLING ONLY of bulk powder (any surface):
   a. Sterile vacuum filter sock and
   b. Sterile scissors
   c. HEPA Vacuum
   d. Beveled and non-beveled cardboard collection inlet nozzles
   e. Sterile bag (see above labeling instructions)
   f. Non-sterile sealable bag (see above labeling instructions)

8) SWAB SAMPLING ONLY (irregular surfaces):
   a. Sterile swab
   b. Sterile primary container (plastic swab tube, screw top; remove plastic to label)
   c. Non-sterile self-sealing bag (see above labeling instructions)

9) LIQUID SAMPLING ONLY
   a. Sterile transfer pipette and sterile swab(s)
   b. Sterile primary container (plastic tube/wide mouth cup; see above labeling instructions)
   c. Non-sterile self-sealing bag(s) (see above labeling instructions)
   d. Parafilm
**SAMPLING TOOLS AND SUPPLIES**

**Sampling Tools** – Sterile tools for collecting product.

- Sterile Sampling Card
- Sterile Macrofoam Swab
- Sterile Vacuum Sock
- Sterile Transfer Pipette

**Primary Containers** – sterile containers for depositing collected product.

- Sterile Collection Cup
- Sterile Baggie (2 sizes)
- Sterile Collection Tube

**Secondary containers** – non-sterile containers for depositing primary containers.

- Small Non-Sterile Sealable Baggie
- Large Non-Sterile Sealable Baggie
- Large Non-Sterile Pouch

**Other** – Supportive materials for sampling.

- Parafilm
- Cardboard Vac Tubes
**PRIMARY SOURCES and BULK POWDERS METHOD**

1) *Assistant Sampler*- Stage away from sampling area.

2) *Assistant Sampler*- If the source of the powder is a letter or small package, open the pre-labeled PRIMARY SOURCE bag, positioning it above the surface next to the source (*if source is larger than W24”xD16”xH10” obtain prior approval from PHL before submitting)*.

3) *Sampler*- Gently place the SOURCE in the bag so “SOURCE markings” are visible through bag. Remove outer gloves and place in discard bag.

4) *Assistant Sampler*- Seal PRIMARY SOURCE bag and place in its secondary bag also labeled PRIMARY SOURCE. Set this bag aside for decontamination.

5) *Assistant Sampler*- Open the sterile laminated collection card pouch by separating clear plastic from its paper backing.

6) *Sampler*- Remove the collection card.

7) *Assistant Sampler*- Open the sterile swab packaging at end opposite of swab head.

8) *Sampler*- Remove swab. Hold the laminated card at an angle to the surface, next to the powder. Use the card to gently push the powder into a pile. With slow deliberate motions use the swab to gently move the powder onto the laminated card.
9) **Assistant Sampler**- Uncap sterile container labeled BULK POWDER and firmly hold the container horizontally, near the sampling area.

10) **Sampler**- Slowly place the laminated card into the BULK POWDER container firmly held by **Assistant Sampler**.

11) **Assistant Sampler**- Secure the BULK POWDER container by slowly screwing its cap in place, being careful not to cross-thread. Set container aside.

12) **Assistant Sampler**- Uncap sterile tube labeled DRY SWAB for **Sampler** to deposit used swab.

13) **Sampler**- Deposit swab into tube held by **Assistant Sampler**. Avoid contact with outside of the tube.

14) **Assistant Sampler**- Recap DRY SWAB tube without cross-threading. Place this tube into bag labeled DRY SWAB. Seal bag and set aside for decontamination. Open BULK POWDER bag.

15) **Sampler**- Without touching the bag, place the sealed container of bulk powder into BULK POWDER bag held open by the **Assistant Sampler**. Remove and discard outer pair of gloves.

16) **Assistant Sampler**- Seal BULK POWDER bag and decontaminate outside of sample bags with a 10% bleach solution. Remove and discard outer pair of gloves. Proceed to decon with samples.
1) **Assistant Sampler** - Stage away from sampling area.

2) **Assistant Sampler** - Assemble vacuum and plug into power source.

3) **Sampler** - Cut open sterile dust collection sock packaging with sterile scissors and remove collection sock. Discard packaging and scissors in discard bag.

4) **Sampler** - Open sock from top to bottom. Insert the white, closed end of filter sock into the larger of the two cardboard tubes. Pull down the upper blue portion of filter sock to covering outside of cardboard nozzle.
5) Snugly insert the smaller cardboard nozzle (non-beveled end) into the open filter sock.

6) **Sampler** - Snugly insert open end of the larger cardboard tube onto vacuum hose. Turn on the vacuum and slowly make a pass of the sampling area.

7) **Sampler** - Turn off the vacuum and remove cardboard nozzle from the vacuum hose. **DO NOT take apart sampling tubes.** Deposit assembled sampling tubes into primary container (pre-labeled, sterile bag) held open by **Assistant Sampler.**
SWABBING IRREGULAR SURFACES METHOD

1) **Assistant Sampler**- Stage away from sampling area.

2) **Assistant Sampler**- Open sterile swab package.

3) **Sampler**- Remove swab by its shaft.

4) **Assistant Sampler**- Uncap the tube labeled **SWAB**.

5) **Sampler**- Swabs surface slowly rolling swab to collect product on all sides of swab.
6) **Sampler** - Place swab in tube held firmly by **Assistant Sampler** taking care not to touch outside of tube with swab.

7) **Assistant Sampler** - Recap and place the tube in the secondary SWAB bag. Seal bag.

8) **Assistant Sampler** - Decontaminate the outside of SWAB bag with a 10% bleach solution.
LIQUID SAMPLING METHOD

1) **Assistant Sampler**- Stage away from sampling area.

2) **Assistant Sampler**- Aseptically open a transfer pipette.

3) **Sampler**- Remove the transfer pipette. Pinch the bulb of the pipette as you insert the pipette tip in the fluid. Release pinch to draw fluid into the pipette (if substance is too thick, apply swab method).

4) **Assistant Sampler**- Uncap the primary container (tube) and hand to **Sampler**.

5) **Sampler**- Insert tip of pipette into collection tube held and pinch the bulb to release liquid (or if swab was used insert swab, tip first).
6) **Sampler**- Recap tube and stretch parafilm around cap and tube securing cap.

7) **Sampler**- Place the sealed sample tube (primary container) into a self-sealing bag (secondary container) held open by the *Assistant Sampler*. Sampler does not touch the outside of the secondary container.
POST-SAMPLING ACTIVITIES

1) Apply tamper tape to each decontaminated secondary container’s seal and write your initials on the tape overlapping onto the bag.

2) Place secondary container(s) in a clean biohazard bag that has remained in the cold zone throughout event. Place biohazard bag into a hard sided container for transport.

3) Complete Sample Collection and Screening Form. Each individual having the specimen in their possession will need to sign on the chain-of-custody section.

4) Call the Public Health Laboratory’s 24/7 call phone (405-406-3511) prior to transport.

**ALL SAMPLES MUST BE SCREENED PRIOR TO ACCEPTANCE BY PHL**